



## **MONTHLY ACTIVITY REPORT**

**January 2018**

# Table of Contents

ABSTRACT..... 1

MAPPING (HOUSEHOLD CONTAINER MONITORING) ..... 2

OVITRAP SURVEILLANCE ..... 3

BG SURVEILLANCE..... 15

RESISTANCE TESTING..... 20

DENGUE ANTIGEN TEST ..... 23

TRAINING ACTIVITIES ..... 25

PRESENTATIONS..... 26

APPENDICES ..... 27

    APPENDIX I Surveillance Map ..... 27

    APPENDIX II Resistance Datasheet ..... 28

    APPENDIX III Resistance Protocol ..... 29

    APPENDIX IV Dengue Test Graphics ..... 30

    APPENDIX V Consent Form ..... 31

    APPENDIX VI Glossary ..... 33

## ABSTRACT

The Belize Vector and Ecology Center has embarked on vector surveillance initiatives that will allow us to determine changes in distribution and density of vectors and obtain information about population changes. In turn this will allow us to monitor and evaluate adequate control methods. In this report, we have outlined the various activities that we have been conducting for the month of January.

The **Household Container Mapping** is being conducted by teams consisting of BVEC employees and Vector Control representatives. So far we have mapped approximately 17.73% of the total structures in Orange Walk Town. Mapping data was updated for the month of January based on the amount of structures we did.

**Ovitrap Surveillance** is being done in which 120 oviposition cups are set out to 60 pre-consented homes in Orange Walk Town. The hatch rate has been determined to be about 40 to 60% and there is interesting variation in the number of eggs collected per zone which may also be related to the position of ovicups and climatic conditions.

The **BG Sentinel Surveillance** is being conducted in order to collect live adult *Aedes* mosquitoes in the field. There are 12 BG Sentinels in the town at 12 pre-consented homes. The traps so far have collected a number of *Aedes* mosquitoes as well as some from the *Culex* species and other types of flies.

The adult *Aedes* mosquitoes collected in the traps are then sorted, identified and stored for testing using the **Dengue Antigen Kit**. Tests conducted so far have turned up negative which is a good sign and will continue testing while working on a pooling strategy depending on the number of adult females collected in the field.

**Larval Resistance Testing** is another key component to the surveillance which will be done in conjunction with the ovitrap surveillance initiative. Currently eggs are being pooled and hatched so that there is enough larvae to meet the standard testing methods. For this month some resistance curves were established for some of the zones as the contamination issues were addressed.

BVEC also does **Presentations** and **Training** sessions in which we collaborate with schools and other groups to foster community engagement.

## MAPPING (HOUSEHOLD CONTAINER MONITORING)

The MOH OW Vector Control Department has been undergoing an initiative in which we map the structures within Orange Walk Town by acquiring GPS data points and gathering structure information (i.e. level of structure, type, no. of doors, windows etc). This data is being linked to the daily *Aedes aegypti* yard inspection that the MOH is collecting. All the data is being collected with the use of an online form that is displayed on a tablet and utilized by BVEC research assistants. Data collection is done in teams with the presence of at least one vector control representative. Each structure is coded with a unique structure ID that is generated based on the zone, street name and number of the structure. The zonation is obtained from a previous database that the town council used in order to facilitate proper sanitation of the town. Orange Walk town is divided into 6 Zones labelled from A to F. The Ministry of Health uses these same zones so that each Vector Control representative has their designated zones that they are assigned to. Below are data collected so far that has been analysed up to the month of January 2018.

Structures mapped	Total
House	846
House/store	34
Store	13
Apartment/Hotel	10
Lab	0
Church	8
Bar	2
House/bar	2
Restaurant	1
School	4
Cemetery	1
Office	3
Clinic	1
Business	13
Other	5
<b>TOTAL STRUCTURES</b>	<b>5420</b>
<b>TOTAL MAPPED</b>	<b>961</b>
<b>PERCENTAGE COMPLETED</b>	<b>17.73%</b>

Aedes Postive Containers	Zone A	ZoneB	Zone C	Zone D	Zone E	Zone F	Total
Vat/tank	3	0	0	0	0	1	4
Drum	8	2	1	0	4	5	20
Bucket	1	2	3	0	5	1	12
Bottle/cans	0	0	0	0	0	0	0
Tires	2	3	1	0	15	2	23
Treeholes/wells	0	0	0	0	0	0	0
Other	5	6	5	0	11	3	30
<b>Grand Total</b>	<b>19</b>	<b>13</b>	<b>10</b>	<b>0</b>	<b>35</b>	<b>12</b>	<b>89</b>

Table 2. Aedes positive containers per zone

Common Containers Types	Zone A	Zone B	Zone C	Zone D	Zone E	Zone F	Total
Vat/tank	21	21	8	0	16	9	75
Drum	32	8	6	0	15	16	77
Bucket	71	57	25	0	82	47	282
Bottle/cans	125	172	142	0	260	91	790
Tires	30	10	4	0	61	13	118
Treeholes/wells	9	0	0	0	2	8	19
Other	210	171	70	0	164	188	803
<b>Grand Total</b>	<b>498</b>	<b>439</b>	<b>255</b>	<b>0</b>	<b>600</b>	<b>372</b>	<b>2164</b>

Table 3. Number of different types of containers found per zone

Table 1. Types of structures mapped along with percentage completion

**Note: Map displaying GPS points of structures mapped as of January 2018 is attached in the appendices on Page 27**

## OVITRAP SURVEILLANCE

The Belize Vector and Ecology Center is currently undergoing an ovitrap/ovicup surveillance program for six (6) months in Orange Walk Town to capture the eggs of *Ae. aegypti* and *Ae. albopictus*, key vectors in the transmission of Dengue, Zika, and Chikungunya. Homes were selected via a random selection process generated by QGIS software. A total of sixty (60) homes were selected throughout the six (6) zones in Orange Walk Town. Two (2) ovicups/ovitrap were placed in each home, giving a total of 120 ovicups/ovitrap placed throughout the entire town. Note that informed consent was obtained from all home/property owners to participate in the study under the supervision of a Ministry of Health (MoH) Vector Control Officer.

Ovitrap/ovicups are left in the field for a total of seven (7) days, after which the egg papers inside every ovitrap/ovicup is retrieved and taken to the BVEC insectary. Egg papers are carefully folded and placed in a plastic container, egg papers from each home remain separate from one another. The ovitrap/ovicups are cleaned with a scouring sponge and a new labelled egg paper and fresh water are added. Once in the BVEC insectary, the freshly collected egg papers are hung for a 24 hour period to remove any excess water. Once dry, the eggs are counted and the eggs are stored. Note that egg hatching for each zone is being done by pooling eggs from previous weeks, mainly to ensure that there are enough eggs to yield the number of larvae needed to perform a larvicide resistance test. Below is an actual picture of an Ovicup with egg paper in the field.



Figure 1. Oviposition cup with egg paper inside

**Note: A copy of the consent form that was used is attached in the appendices on Page 31**

When hatching eggs, egg papers for a particular zone are carefully inspected for eggs, and then segments of the egg paper containing the eggs are cut and placed in a container with water. The container with eggs is then placed in the vacuum hatcher for hatching. Use of the vacuum hatcher ensures a more synchronized hatch and same size larvae. After a few days, third and early fourth stage larvae are sorted for a larvicide resistance test.

Table 4 below shows the number of eggs collected for each zone for every week. Note that each zone is different in terms of geographic location within the town and whether it is more of a commercial or housing area. The different weather patterns experienced each week are also an important factor influencing oviposition each week. For this reason, the egg density will vary across zones and time.

ZONE	WEEK 13	WEEK 14	WEEK 15	WEEK 16	TOTAL
<b>A</b>	142	169	177	140	<b>628</b>
<b>B</b>	95	151	83	85	<b>414</b>
<b>C</b>	113	56	325	267	<b>761</b>
<b>D</b>	461	265	228	195	<b>1149</b>
<b>E</b>	837	680	664	781	<b>2962</b>
<b>F</b>	58	113	105	165	<b>441</b>
<b>TOTAL</b>	<b>1706</b>	<b>1434</b>	<b>1582</b>	<b>1633</b>	<b>6355</b>

Table 4. Number of eggs collected in each zone per week of surveillance for a total of four (4) weeks surveillance period. Zone E has the highest number of eggs for the entire surveillance period of four (4) weeks with n=2962. Zone B has the lowest number of eggs for the entire surveillance period with n=414.

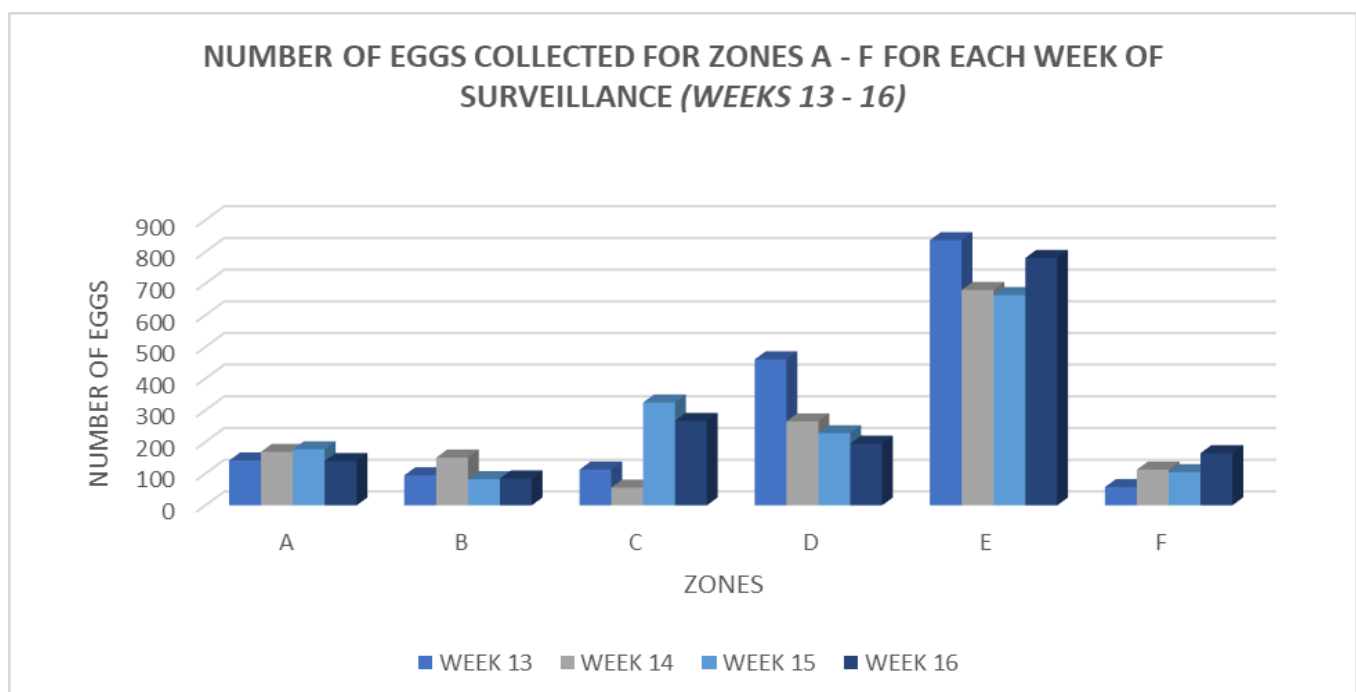


Figure 2. Number of eggs collected per week (4 weeks) for each zone. Zone E has the highest number of eggs collected each week.

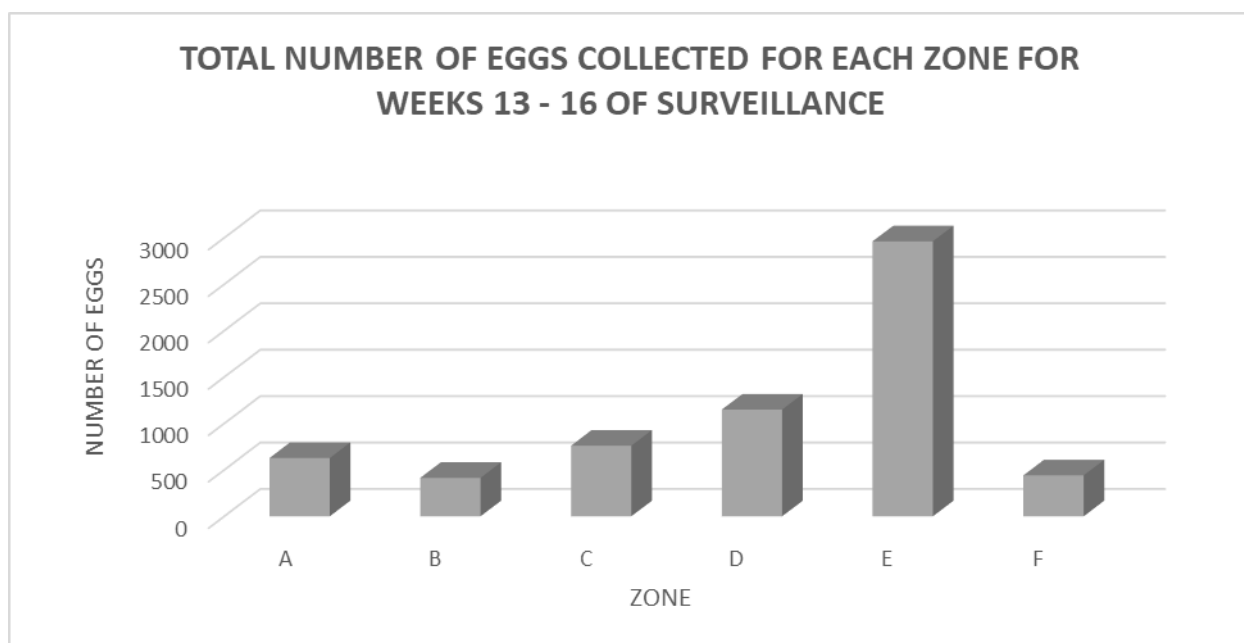


Figure 3. Number of eggs collected in each zone for the entire surveillance period of four (4) weeks/1 month.

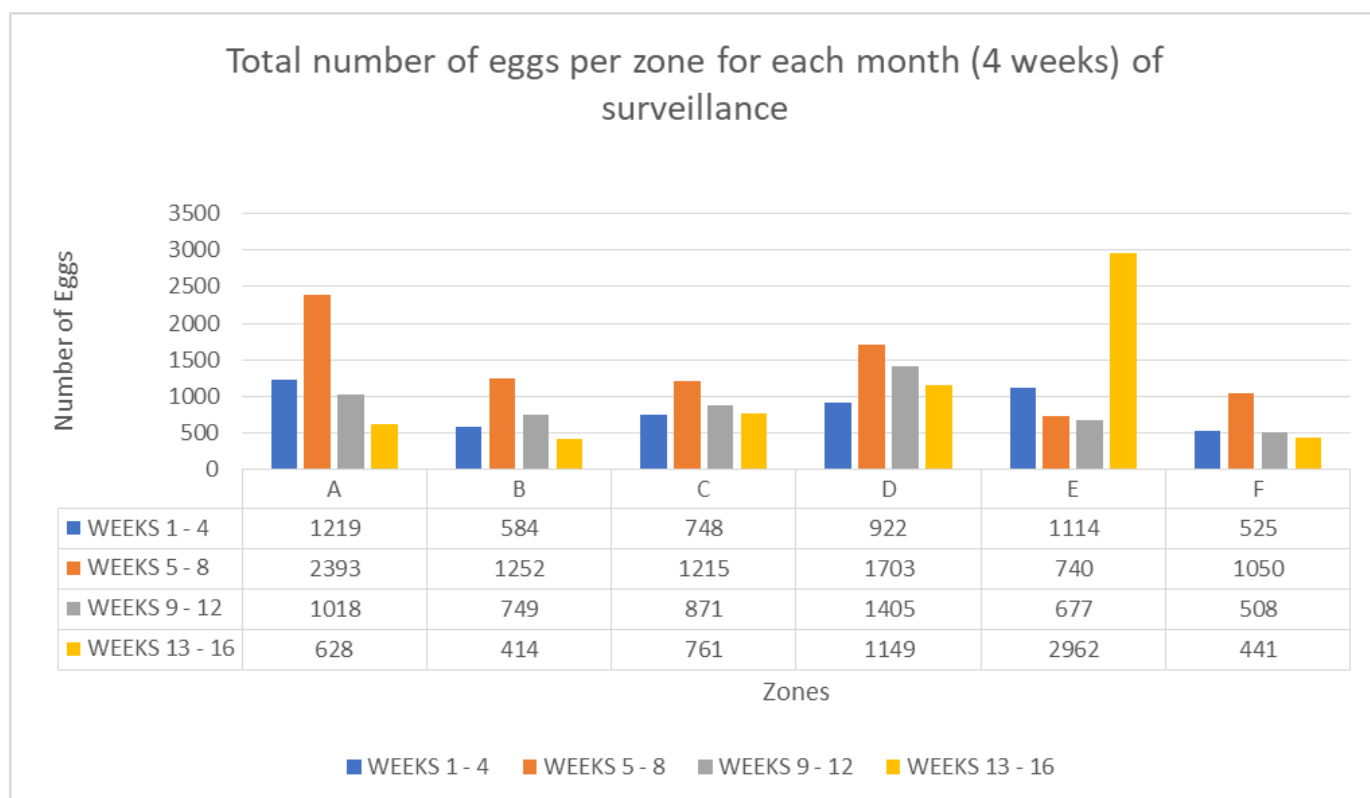


Figure 4. Total cumulative number of eggs collected in each zone for the entire surveillance period (Weeks 1 – 16).

## ZONE A OVITRAPS

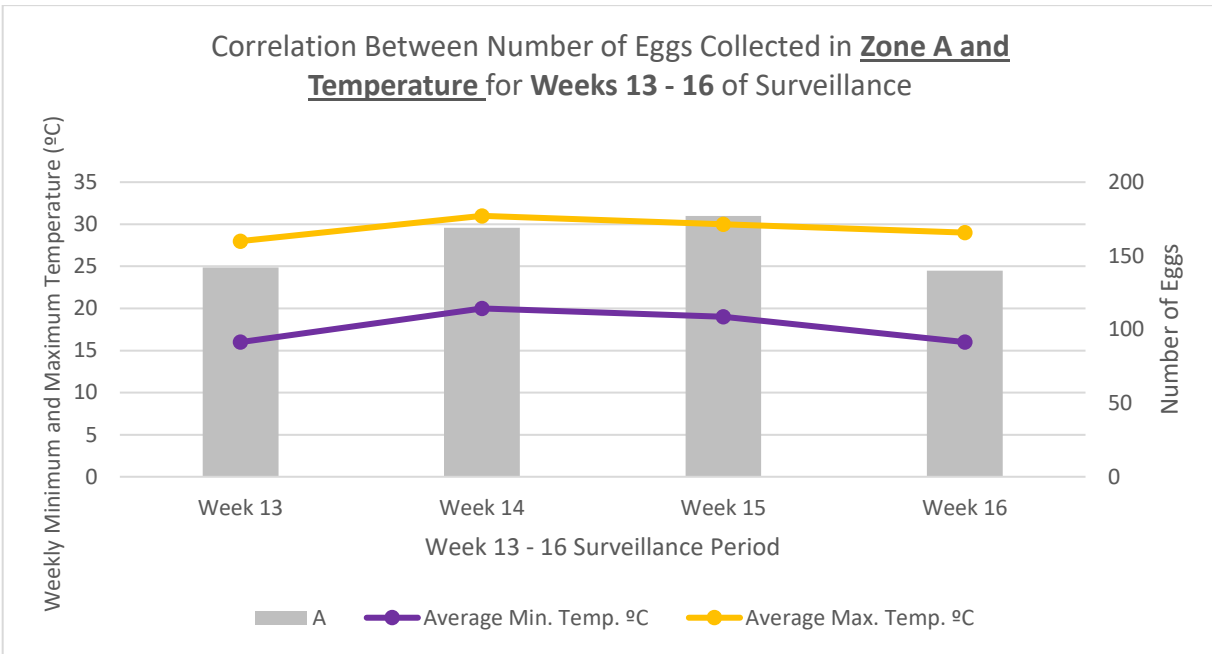


Figure 5. Number of eggs collected in Zone A and temperature data for weeks 13 – 16 of surveillance. Week 15 has the highest number of eggs collected ( $n=177$ ) and Week 16 has the lowest number of eggs collected ( $n=140$ ).

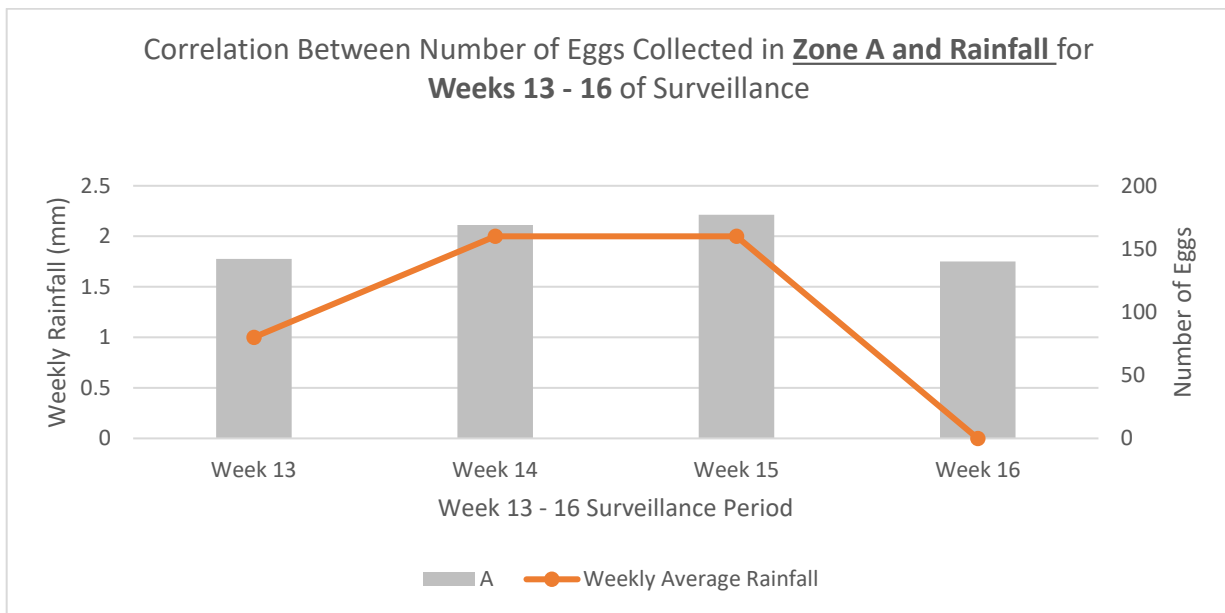


Figure 6. Number of eggs collected in Zone A and rainfall data for weeks 13 – 16 of surveillance.



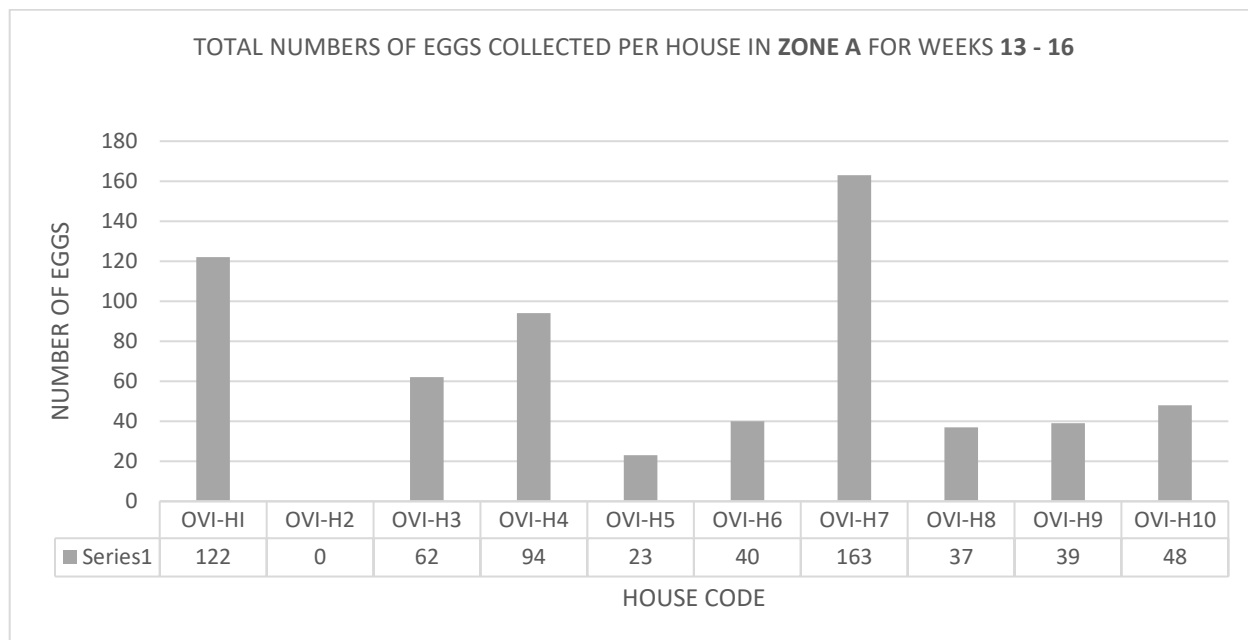


Figure 7. Number of eggs collected per house in Zone A for weeks 13 – 16 of surveillance. House 2 (OVI-H2) has the lowest number of eggs (n=0) and House 7 (OVI-H7) has the highest number of eggs (n=163).

### ZONE B OVITRAPS

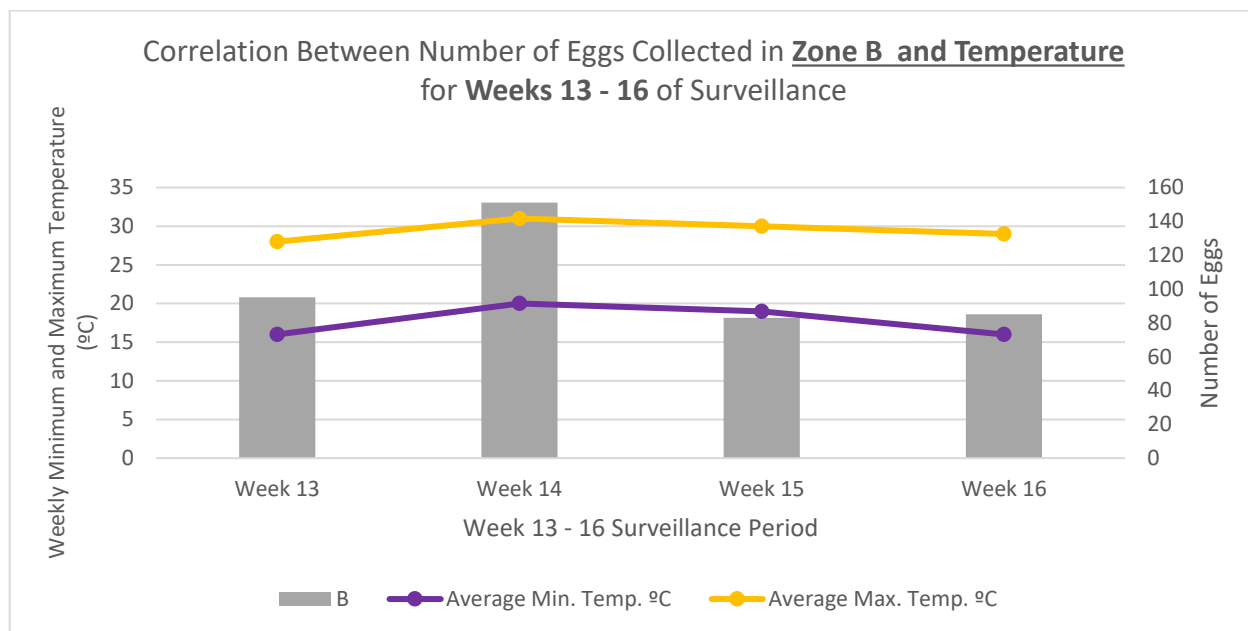


Figure 8. Number of eggs collected in Zone B and temperature data for weeks 13 – 16 of surveillance. Week 14 has the highest number of eggs collected (n=151) and Week 15 has the lowest number of eggs collected (n=83).

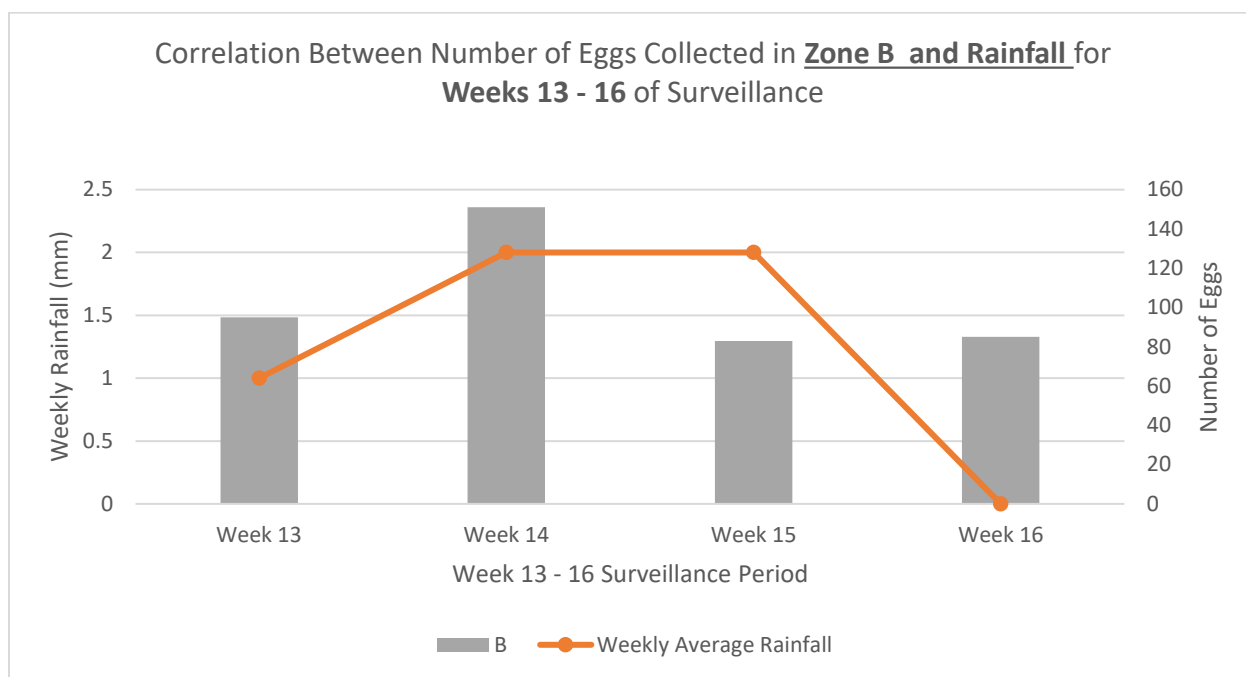


Figure 9. Number of eggs collected in Zone B and rainfall data for weeks 13 – 16 of surveillance.

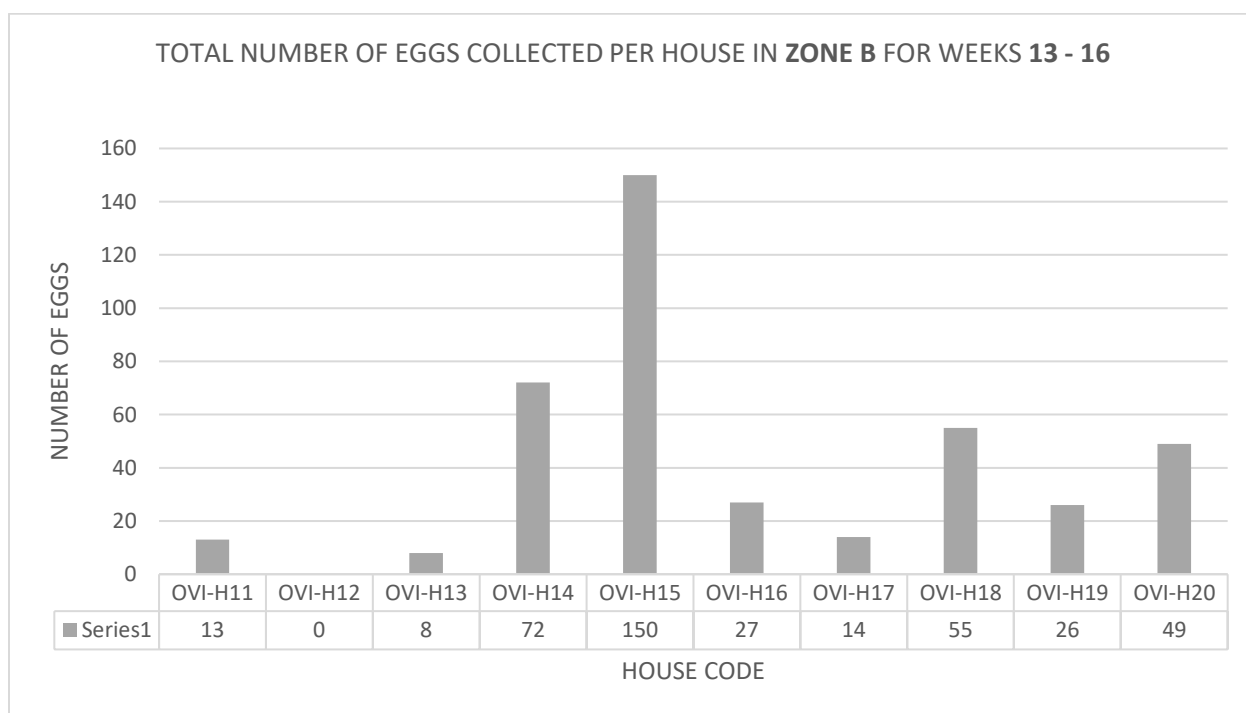


Figure 10. Number of eggs collected per house in Zone B for weeks 13 – 16 of surveillance. House 12 (OVI-H12) has the lowest number of eggs ( $n=0$ ) and House 15 (OVI-H15) has the highest number of eggs ( $n=150$ ).

## ZONE C OVITRAPS

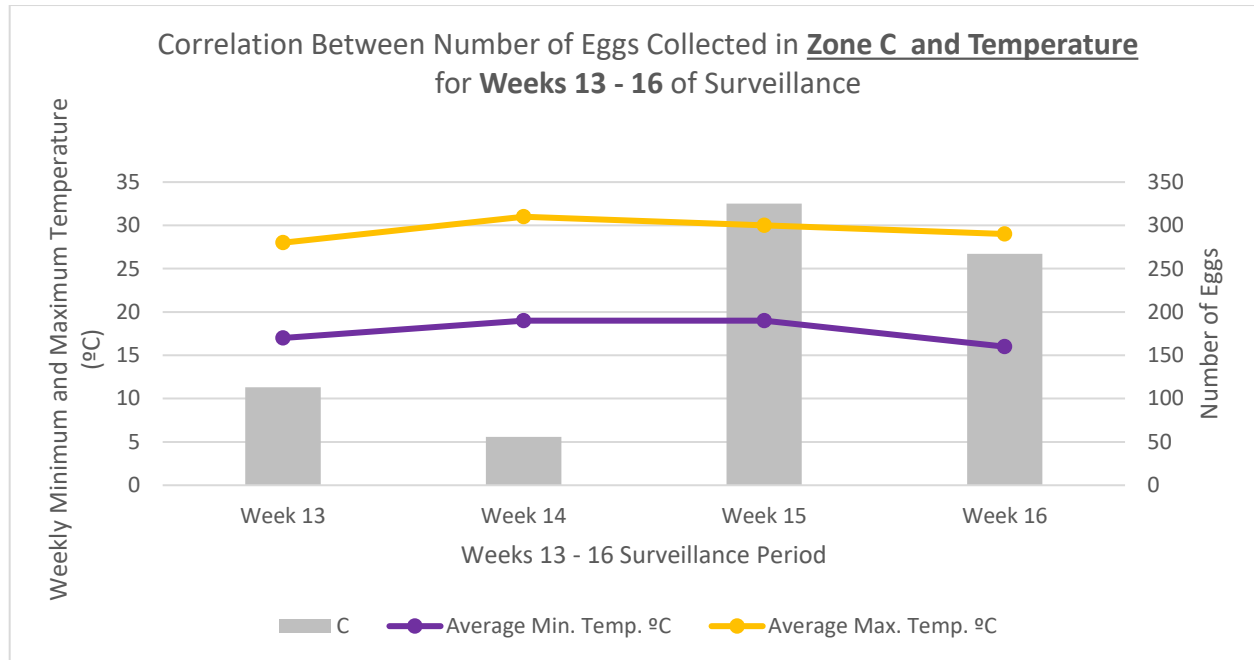


Figure 11. Number of eggs collected in Zone C and temperature data for weeks 13 – 16 of surveillance. Week 15 has the highest number of eggs collected (n=325) and Week 14 has the lowest number of eggs collected (n=56).

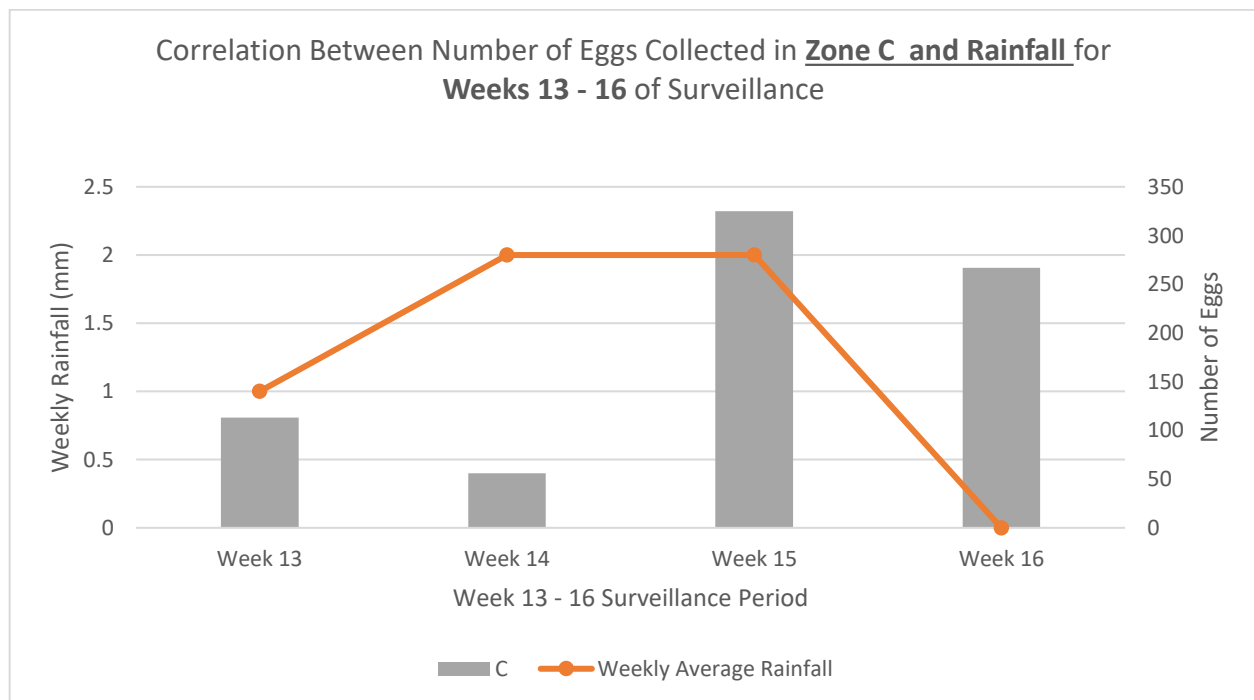


Figure 12. Number of eggs collected in Zone C and rainfall data for weeks 13 – 16 of surveillance.

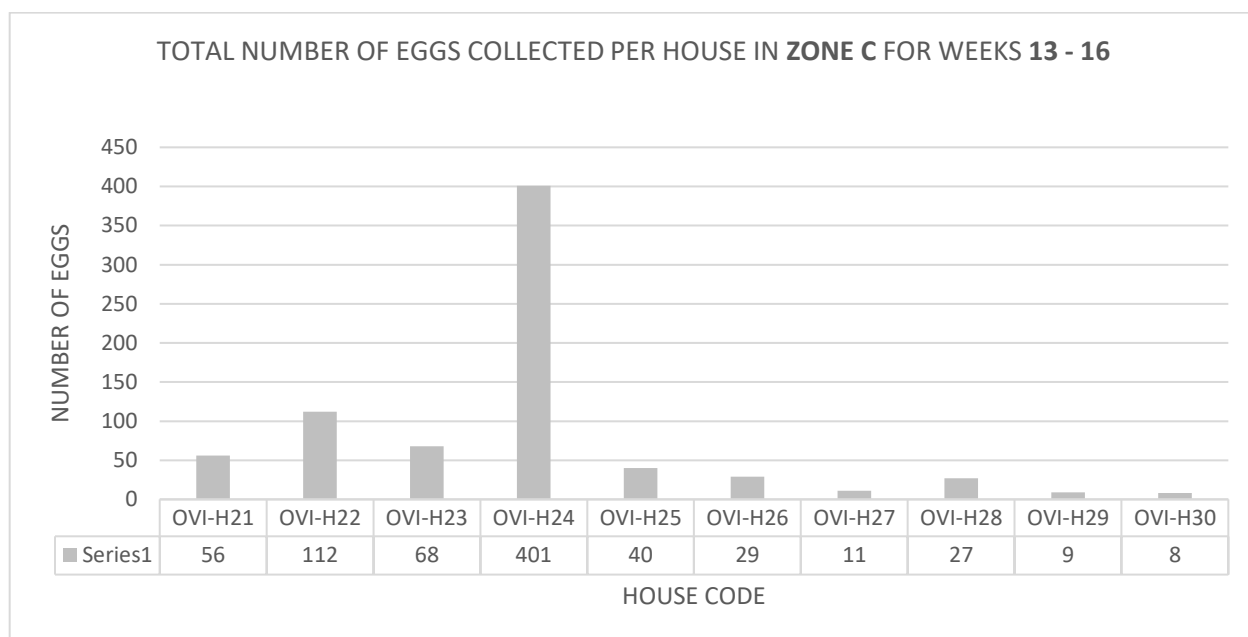


Figure 13. Number of eggs collected per house in Zone C for weeks 13 – 16 of surveillance. House 30 (OVI-H30) has the lowest number of eggs (n=8) and House 24 (OVI-H24) has the highest number of eggs (n=401).

### ZONE D OVITRAPS

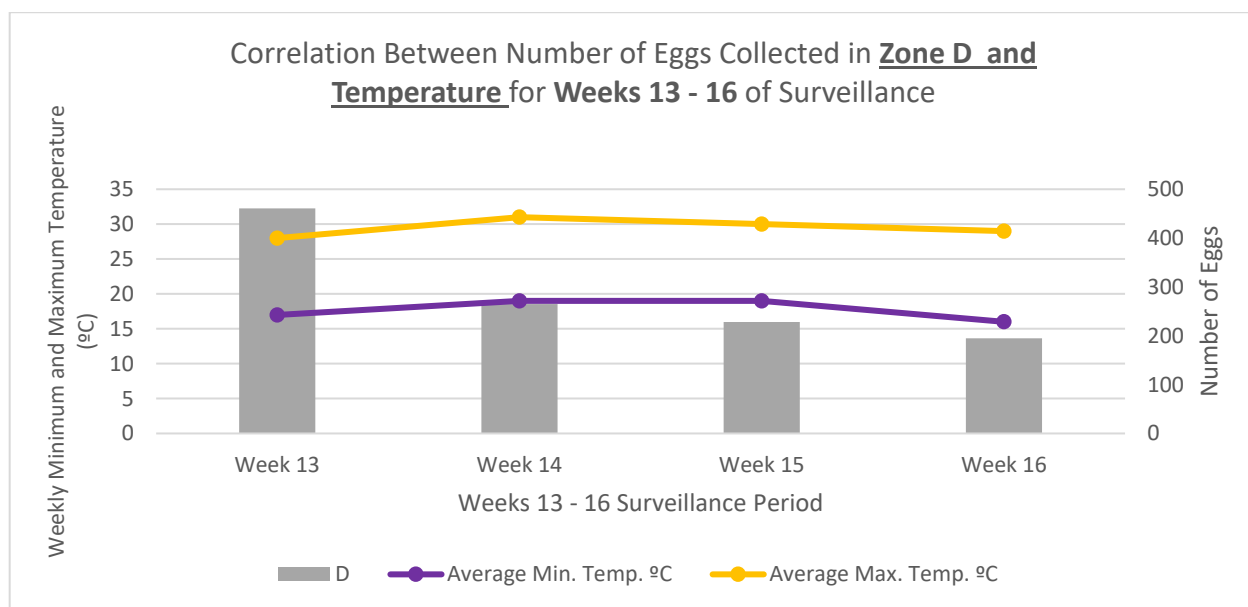


Figure 14. Number of eggs collected in Zone D and temperature data for weeks 13 – 16 of surveillance. Week 13 has the highest number of eggs collected (n=461) and Week 16 has the lowest number of eggs collected (n=195).

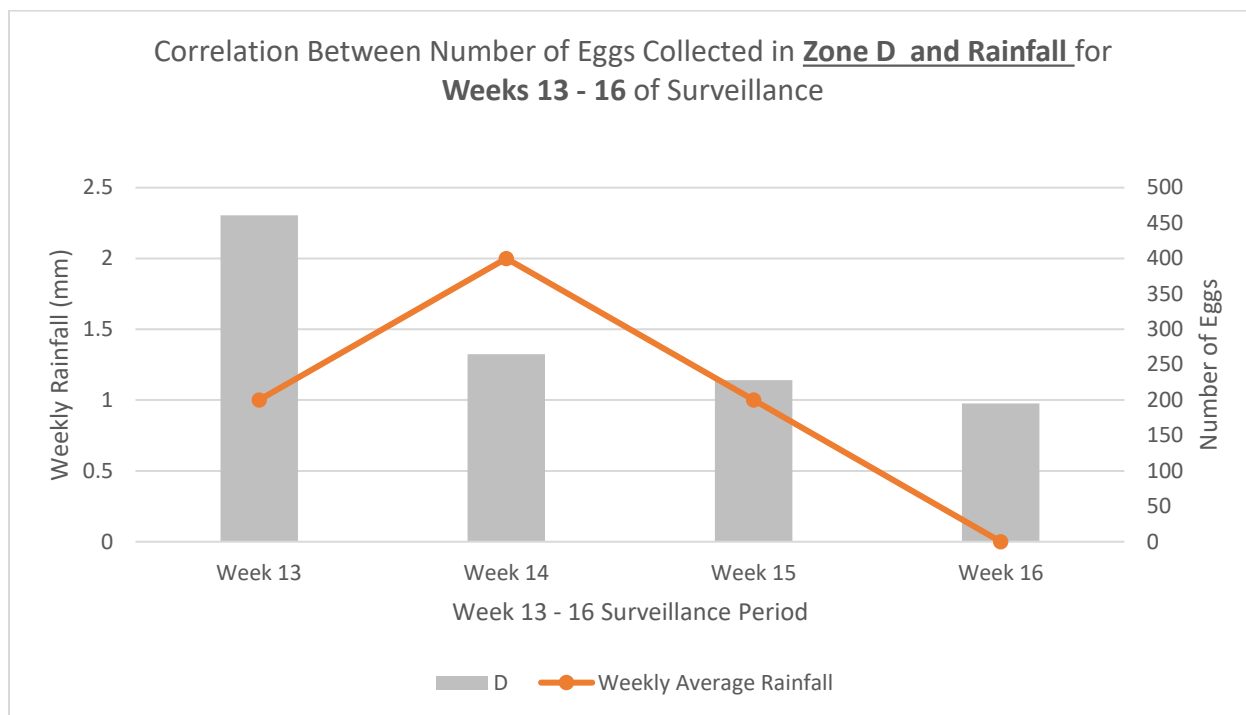


Figure 15. Number of eggs collected in Zone D and rainfall data for weeks 13 – 16 of surveillance.

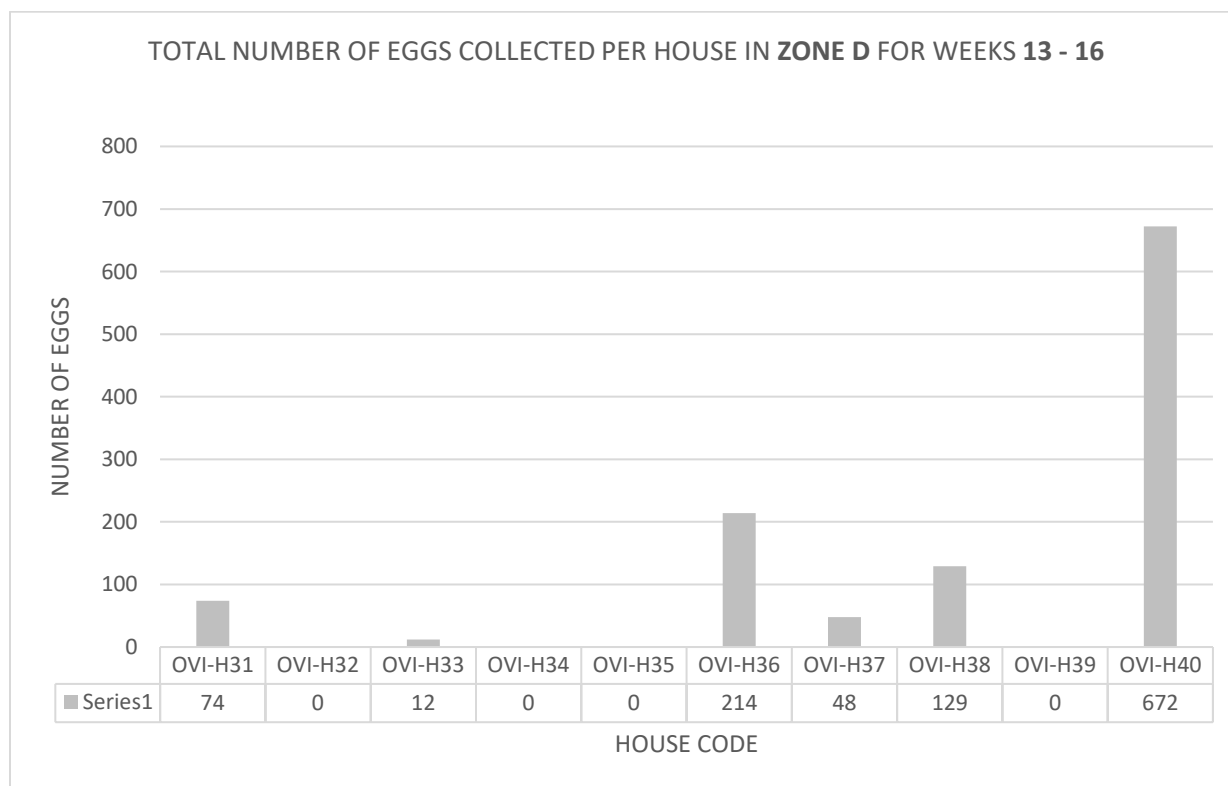


Figure 16. Number of eggs collected per house in Zone D for weeks 13 – 16 of surveillance. House 32, 34, 35, 39 (OVI-H32, OVI-H34, OVI-H35, OVI-H39) have the lowest number of eggs ( $n=0$ ) and House 40 (OVI-H40) has the highest number of eggs ( $n=672$ )

## ZONE E OVITRAPS

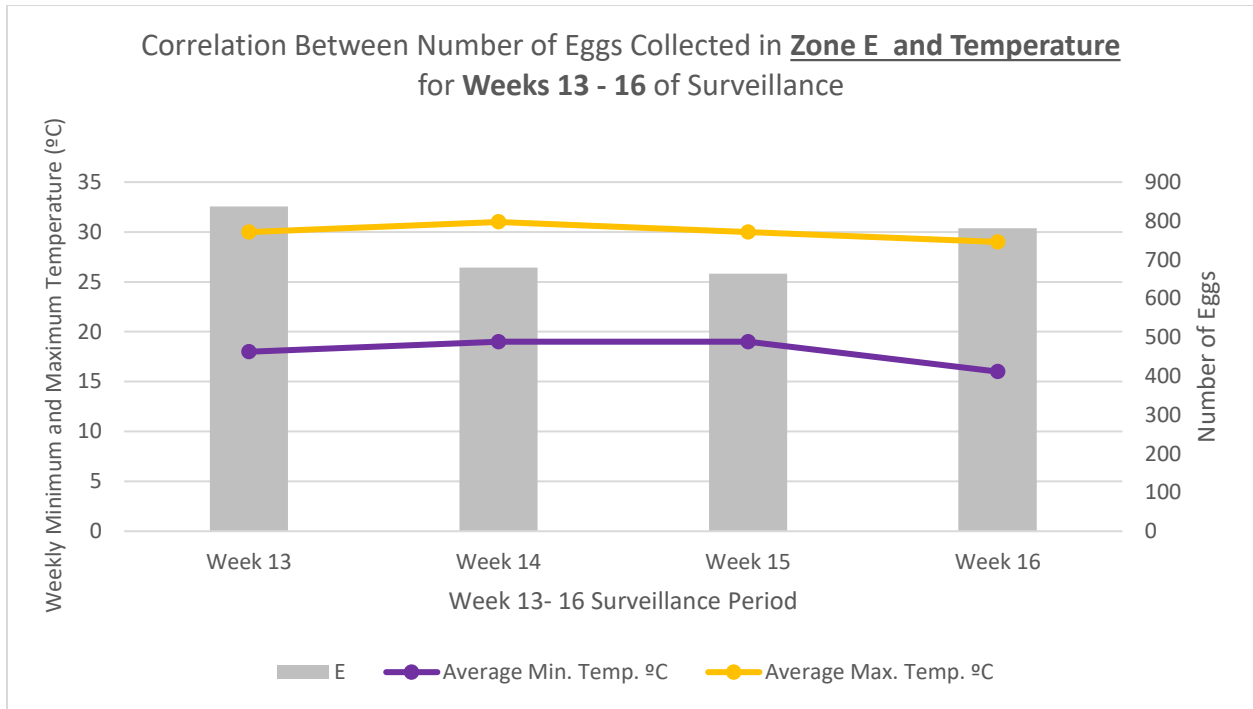


Figure 17. Number of eggs collected in Zone E and temperature data for weeks 13 – 16 of surveillance. Week 13 has the highest number of eggs collected ( $n=837$ ) and Week 15 has the lowest number of eggs collected ( $n=664$ ).

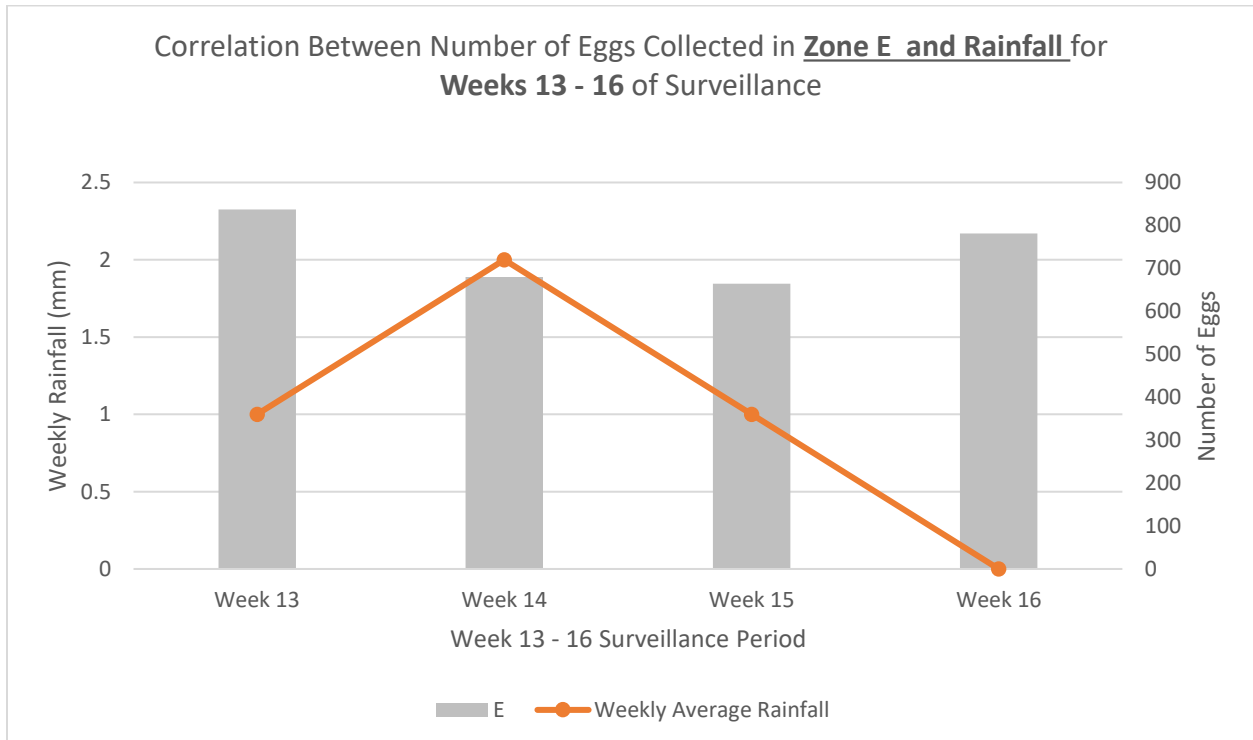


Figure 18. Number of eggs collected in Zone E and rainfall data for weeks 13 – 16 of surveillance.

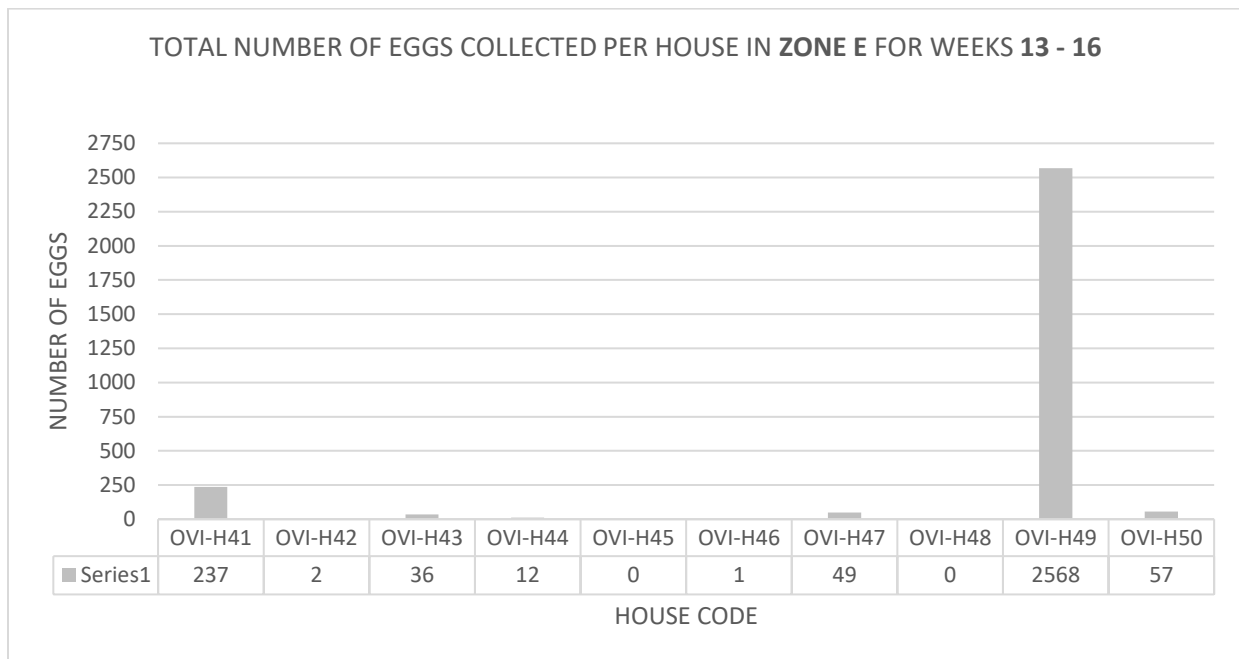


Figure 19. Number of eggs collected per house in Zone E for weeks 13 – 16 of surveillance. House 45 and 48 (OVI-H45 and OVI-H48) have the lowest number of eggs (n=0) and House 49 (OVI-H49) has the highest number of eggs (n=2568).

### ZONE F OVITRAPS

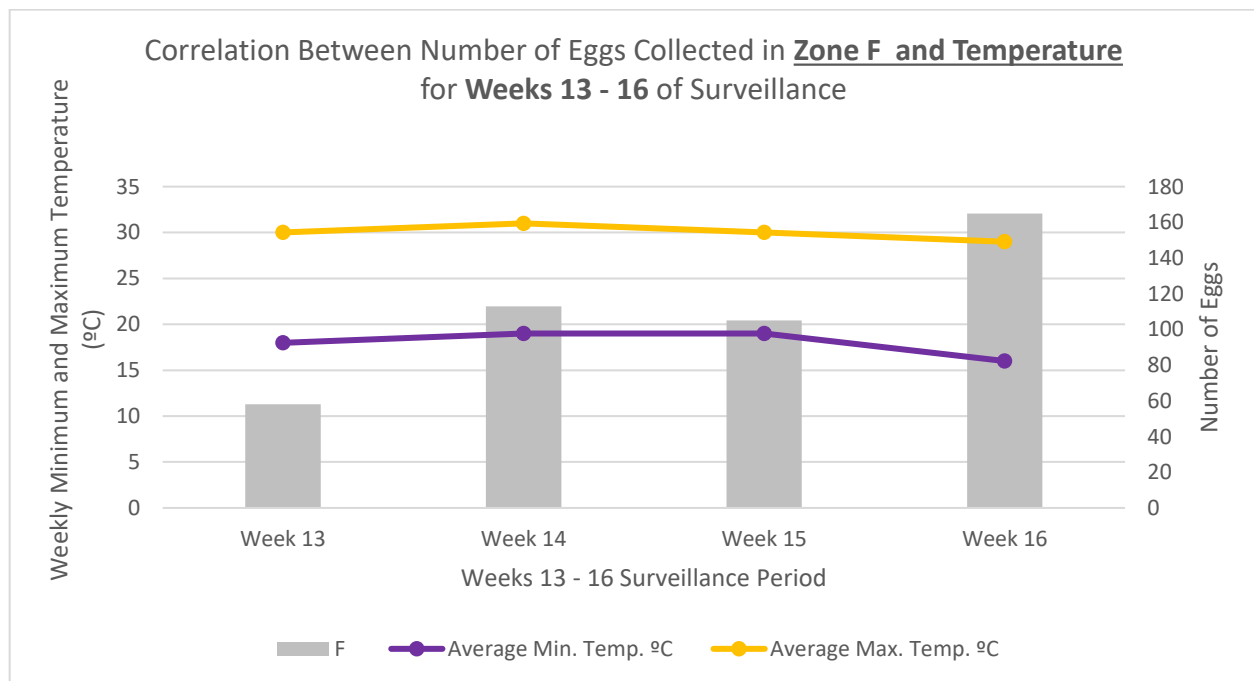


Figure 20. Number of eggs collected in Zone F and temperature data for weeks 13 – 16 of surveillance. Week 16 has the highest number of eggs collected (n=165) and Week 13 has the lowest number of eggs collected (n=58).

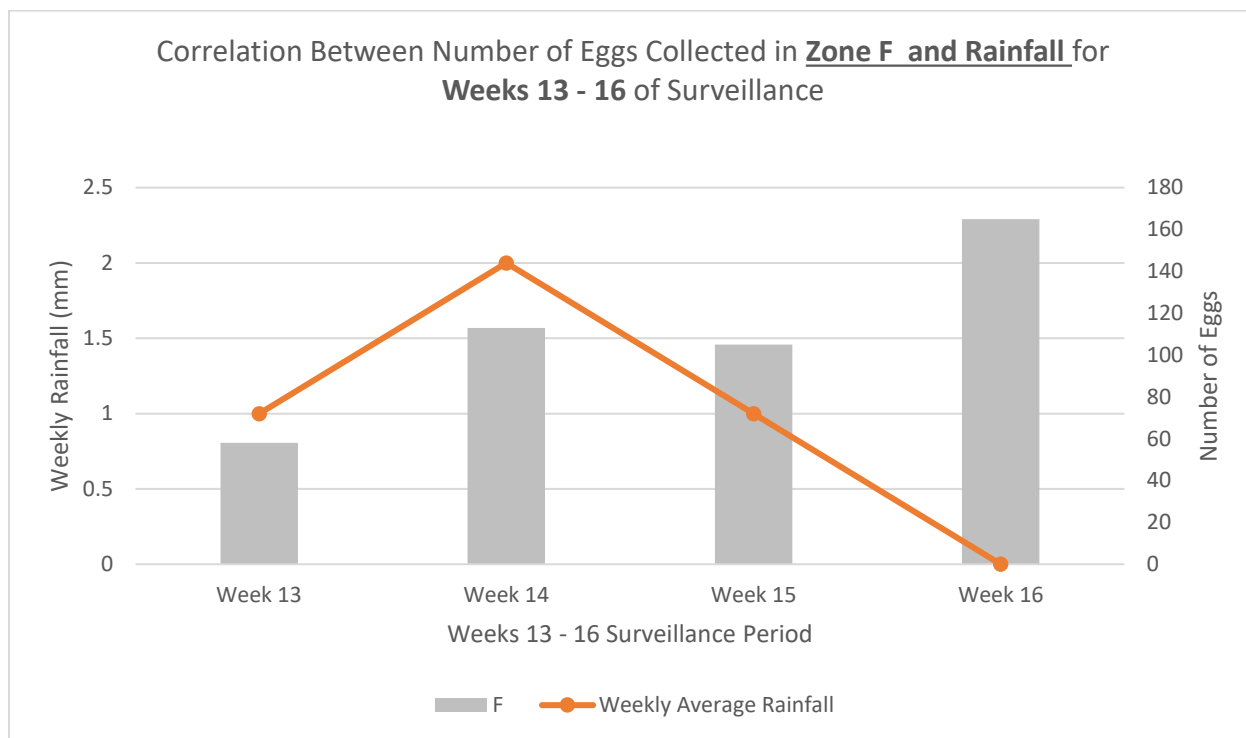


Figure 21. Number of eggs collected in Zone F and rainfall data for weeks 13 – 16 of surveillance.

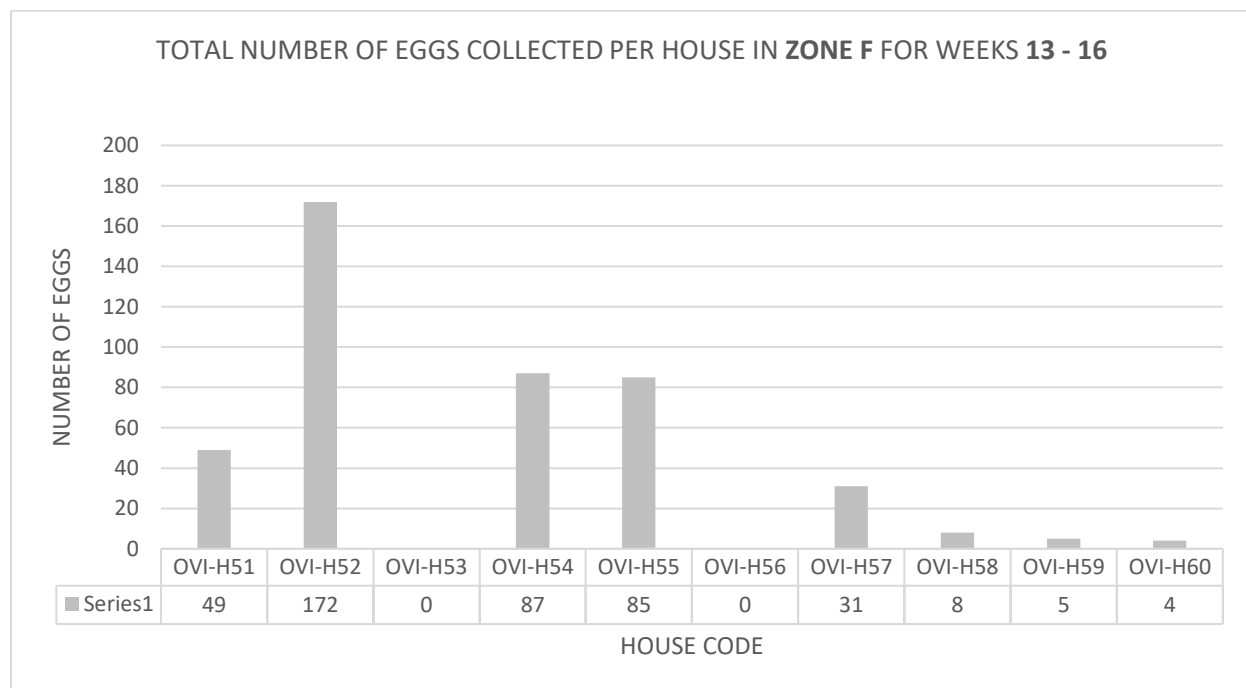


Figure 22. Number of eggs collected per house in Zone F for weeks 13 – 16 of surveillance. House 53 and 56 (OVI-H53 and OVI-H56) have the lowest number of eggs ( $n=0$ ) and House 49 (OVI-H52) has the highest number of eggs ( $n=172$ ).



## BG SURVEILLANCE

For this particular surveillance activity, Biogent (BG) Sentinel traps are used to collect adult *Ae. aegypti* and *Ae. albopictus* mosquitoes in the field. A total of 12 homes (2 homes per zone) were selected via a random selection process generated by QGIS software. Informed consent was obtained from all home/property owners with the supervision of a MOH Vector Control representative.

BGS traps are set out in the field once a week. Note that traps are placed in the mornings and left to operate for 7-8 hours, and then retrieved in the afternoon of the same day. Below is an image of a BG Sentinel taken from one of the houses that is participating in the surveillance initiative.



Figure 23. BG Sentinel trap setup in the field.

The mosquitoes along with other insect specimens collected are taken to the BVEC laboratory and stored for later identification. Identification is done once a week. *Aedes Spp.* and *Culex Spp.* are stored, while any other insect specimens are discarded. Once mosquitoes are identified, they are stored back in the freezer until they can be processed for testing.

ZONE	HOUSE CODE	Total Adult Aedes (♂,♀) Collected Per Week				TOTAL
		Week 13	Week 14	Week 15	Week 16	
		05-Jan-18	12-Jan-18	16-Jan-18	26-Jan-18	
A	BG-H1	0	1	1	0	2
A	BG-H2	1	1	0	0	2
TOTAL		1	2	1	0	4
B	BG-H3	2	1	3	0	6
B	BG-H4	0	0	0	0	0
TOTAL		2	1	3	0	6
C	BG-H5	1	0	5	0	6
C	BG-H6	2	4	6	0	12
TOTAL		3	4	11	0	18
D	BG-H7	4	0	2	0	6
D	BG-H8	0	0	0	0	0
TOTAL		4	0	2	0	6
E	BG-H9	0	1	0	0	1
E	BG-H10	0	0	0	0	0
TOTAL		0	1	0	0	1
F	BG-H11	0	0	0	0	0
F	BG-H12	0	0	1	0	1
TOTAL		0	0	1	0	1
GRAND TOTAL		10	8	18	0	36

Table 5. Number of adult Aedes mosquitoes, both male and female, captured per house in each zone for each week of surveillance. Zone C has the highest number (n=18) of adult Aedes mosquitoes captured while Zone E and F have the lowest number of adult Aedes mosquitoes captured. In terms of weekly captures, week 15 had the highest number (n=18) of mosquitoes captured while week 16 had the lowest number with no mosquitoes captured.

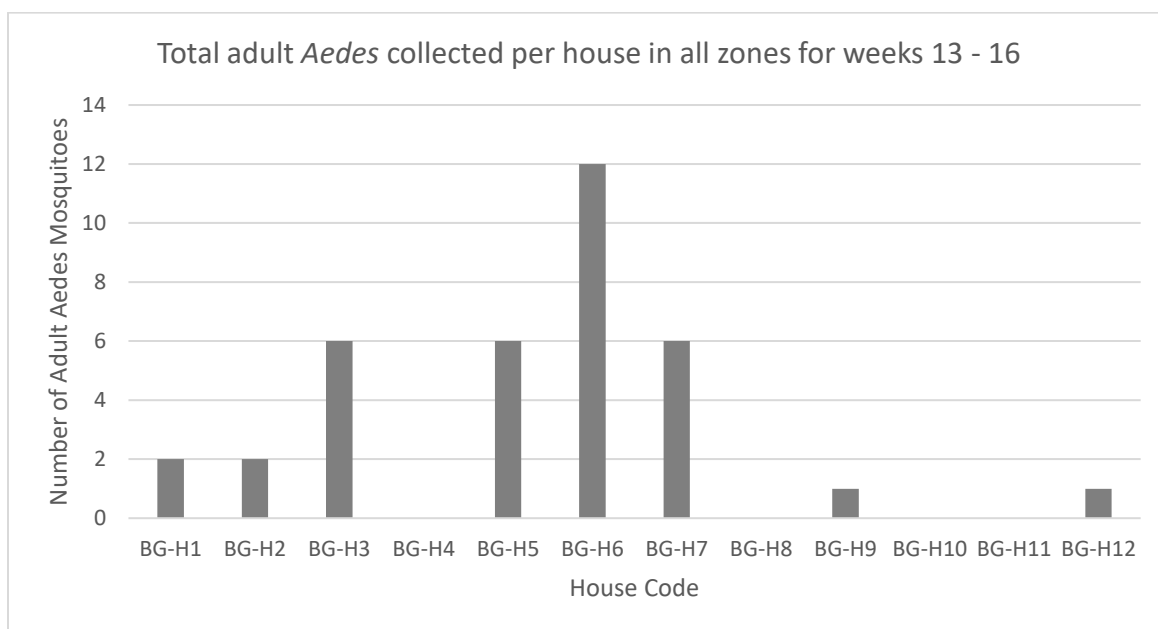


Figure 24. Total number of adult *Aedes* mosquitoes captured per house for all zones for the entire surveillance period of weeks 13-16. House 6 (BG-H6) had the highest number ( $n=12$ ) of adult *Aedes* mosquitoes captured while houses 4, 8, 10 and 11 had no mosquitoes captured.

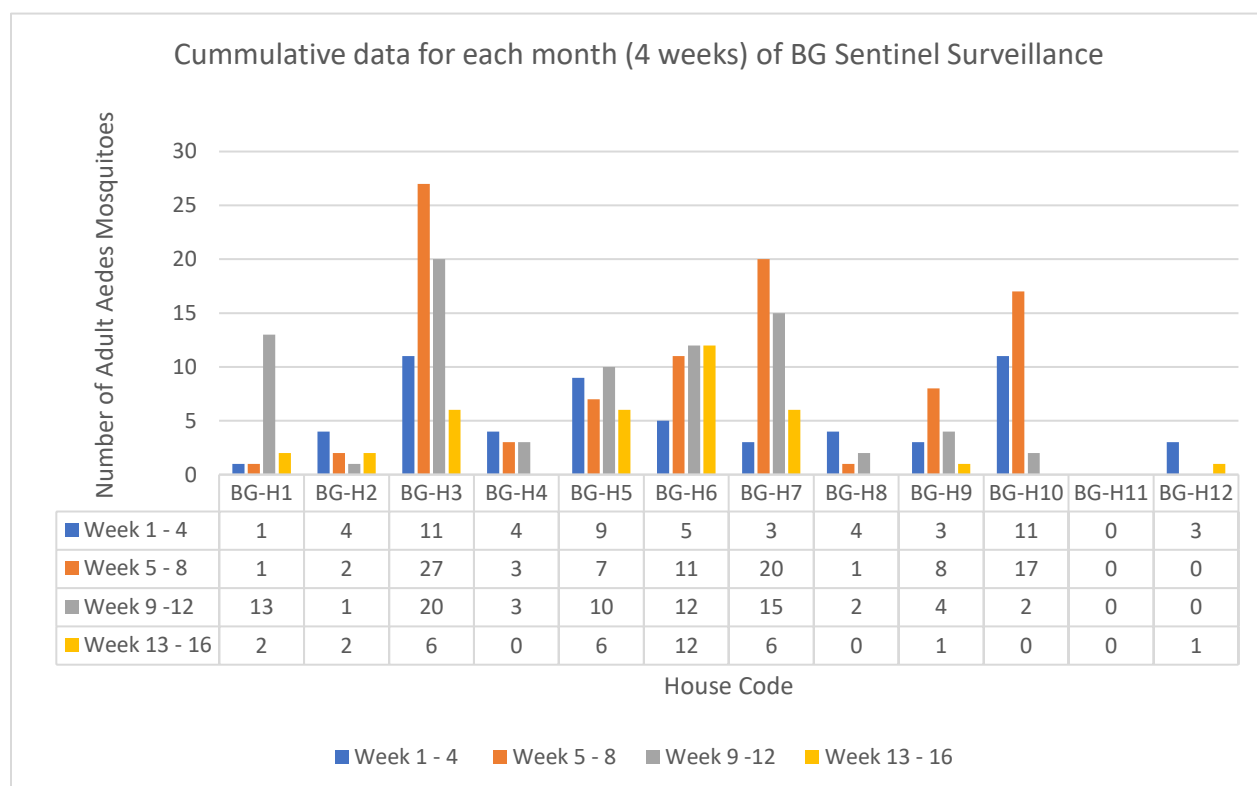


Figure 25. Number of adult *Aedes* mosquitoes captured per house in all zones for each month (4 weeks) of BG Sentinel surveillance thus far.

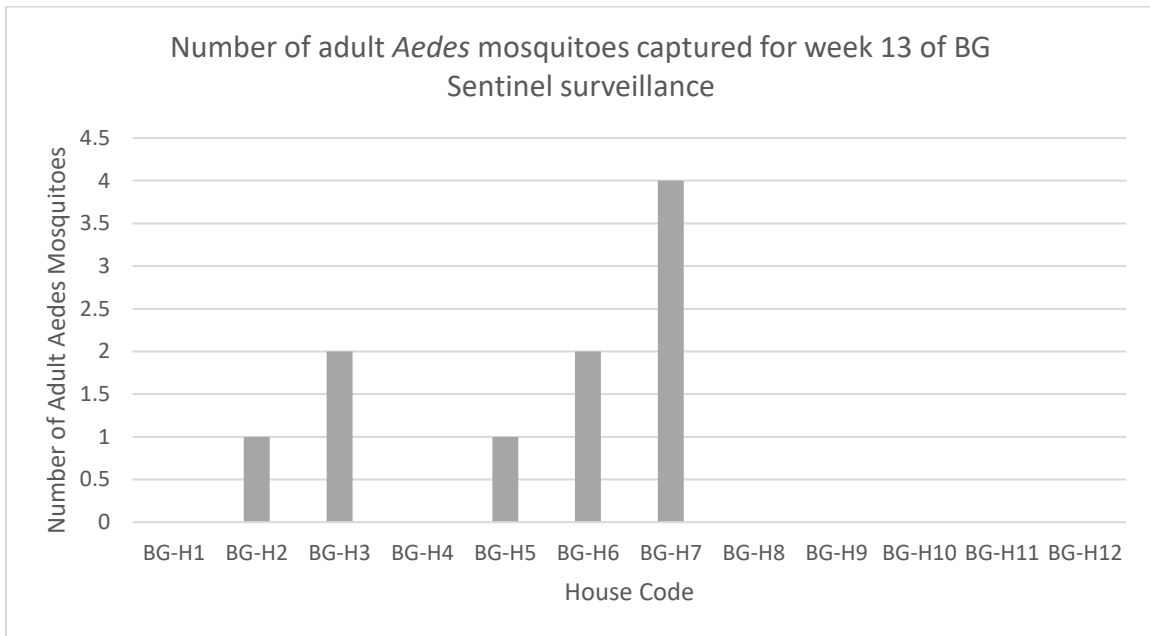


Figure 26. Number of adult *Aedes* mosquitoes captured per house in all zones for week 13 of BG Sentinel surveillance. House 7 (BG-H7) has the highest number of adult *Aedes* mosquitoes captured for that day (n=4).

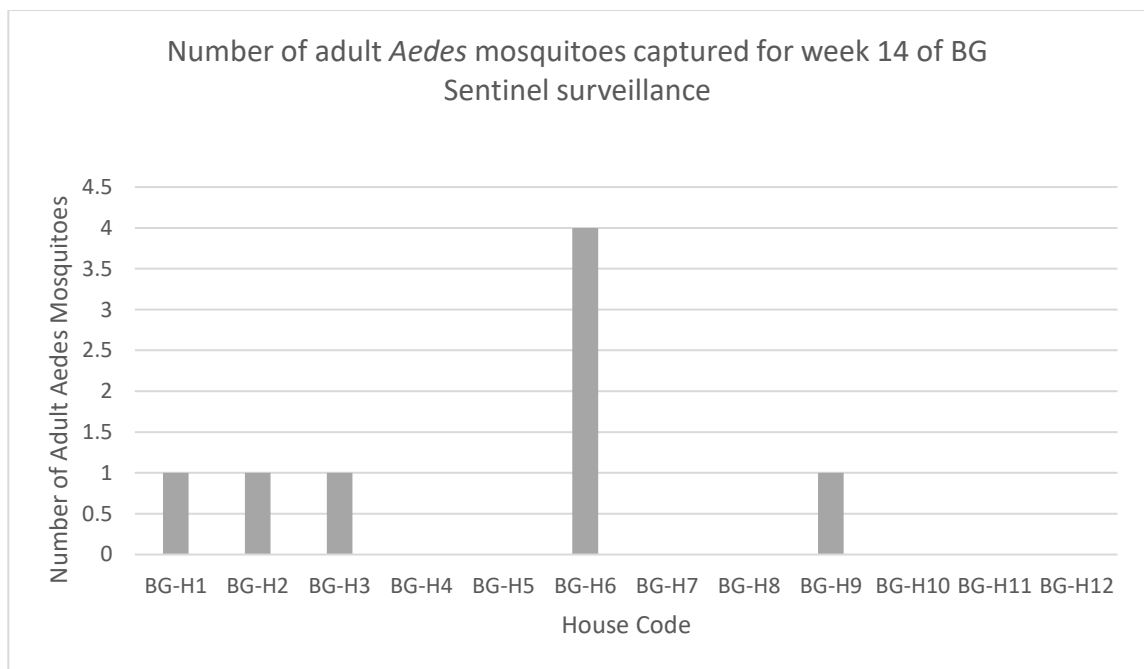


Figure 27. Number of adult *Aedes* mosquitoes captured per house in all zones for week 14 of BG Sentinel surveillance. House 6 (BG-H6) has the highest number of adult *Aedes* mosquitoes captured for that day (n=4).

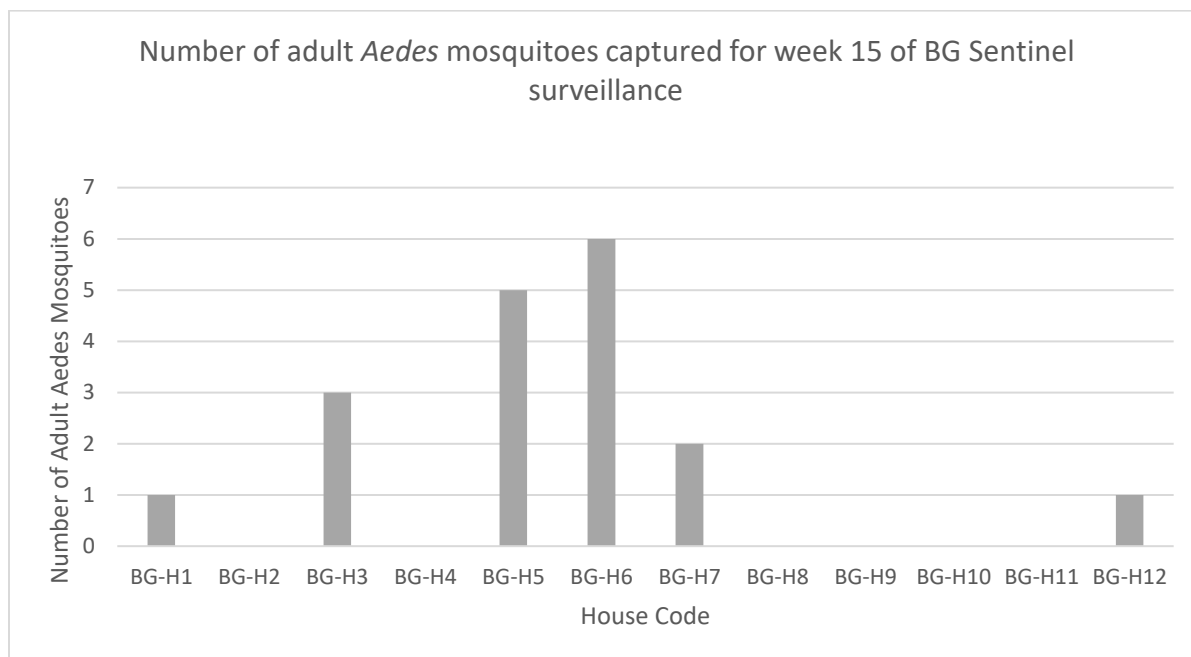


Figure 28. Number of adult *Aedes* mosquitoes captured per house in all zones for week 15 of BG Sentinel surveillance. House 6 (BG-H6) has the highest number of adult *Aedes* mosquitoes captured for that day (n=6).

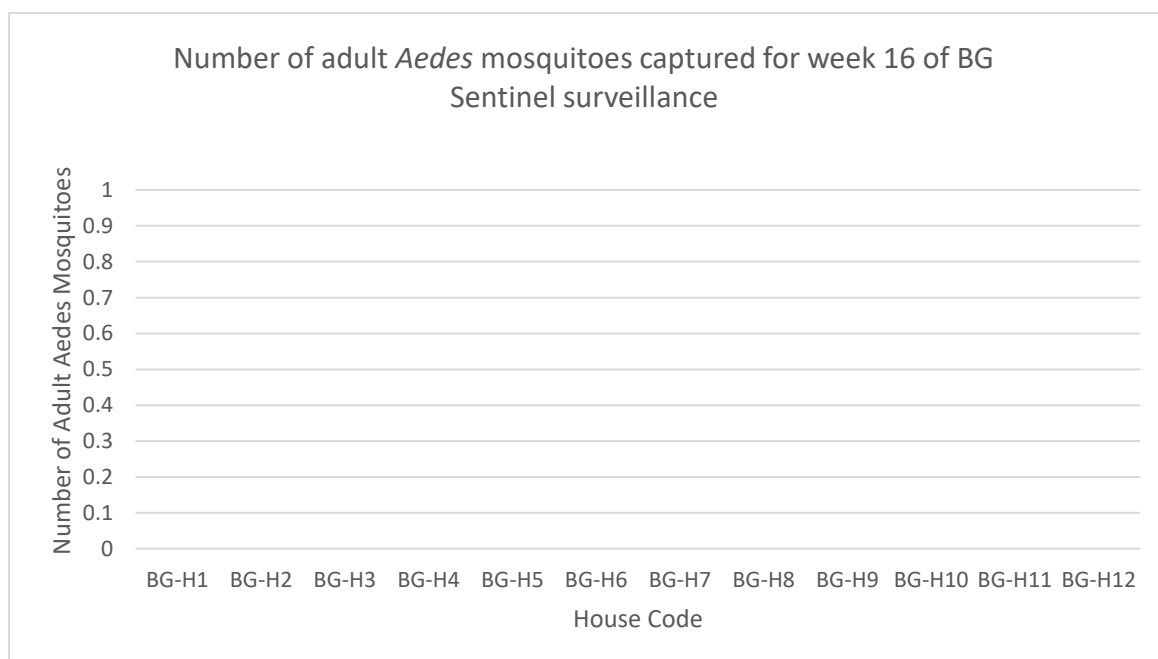


Figure 29. There were no adult *Aedes* mosquitoes captured for Week 16.

## RESISTANCE TESTING

Insecticide resistance testing is being done currently on mosquito larvae that has been reared up from *Aedes* eggs collected via oviposition cups. The following procedure is being conducted based on the World Health Organization standard protocol for determining the susceptibility of mosquito larvae to insecticides. Third and fourth instar larvae are being tested against liquid Temefos (in a solution of 70% ethanol) at varying concentrations. Temefos is the active ingredient found in the granular Abate being used by the Ministry of Health to treat positive larval habitats.

Larvae are reared up in the BVEC insectary based on the zone that they were collected from. After reaching the appropriate size, the larvae are then prepped for testing by separating them into groups of 20 per container. For each trial, 5 containers are used (2 'controls' and 3 'treatments') per concentration per zone which equals to 100 larvae per trial. Containers being used for the test are small glass bowls. Ideally we want to obtain a total of 5 trials per concentration per zone. Depending on the amount of larvae obtained, that will determined how much trials can be done with varying concentrations at a certain time. The 5 glass bowl replicates, are labelled appropriately and are filled with 225ml distilled water. Using a micropipette, 1ml of 70% ethanol is added to both 'control' containers and 1ml of 'x-mg/l Temefos concentration solution' is added to the three 'treatment' containers.

The contents of each container are stirred with a glass stirring rod for 30 seconds to ensure proper mixing. The larvae are then introduced into the containers and the investigator records time introduced, room temperature and relative humidity. These procedures are then repeated for every other trial. The investigator will then check for mortality after 24 hours and after 48 hours while recording any temperature and humidity changes. At the end of the experiment, larvae that have been exposed to the Temefos are then discarded in a large liquid waste disposal container while the control larvae reared up to adulthood and identified thereafter to verify species as either *Aedes aegypti* or *Aedes albopictus*.

The baseline data for Zones F, C and A were established for this month. On the following pages are graphs generated from the QCal software, which we use to calculate the models. Note that specimens from each Zone are tested against a control (ethanol) and 4 different concentrations of Temefos solution (0.005mg/L, 0.025mg/L, 0.125mg/L, and 0.625 mg/L)

## ZONE F, JAN. 24, 2018

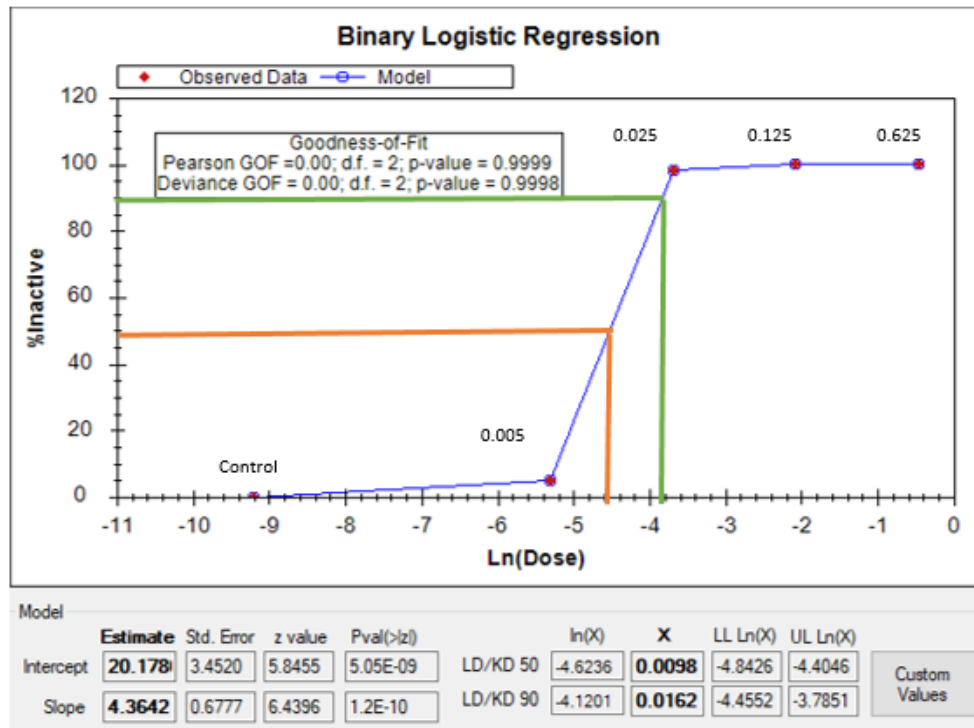


Figure 30. Larvae Resistance Curve for Zone F. LD50 concentration is at 0.0098 mg/L while LD90 concentration is at 0.0162 mg/L

## ZONE C, JAN. 3, 2018

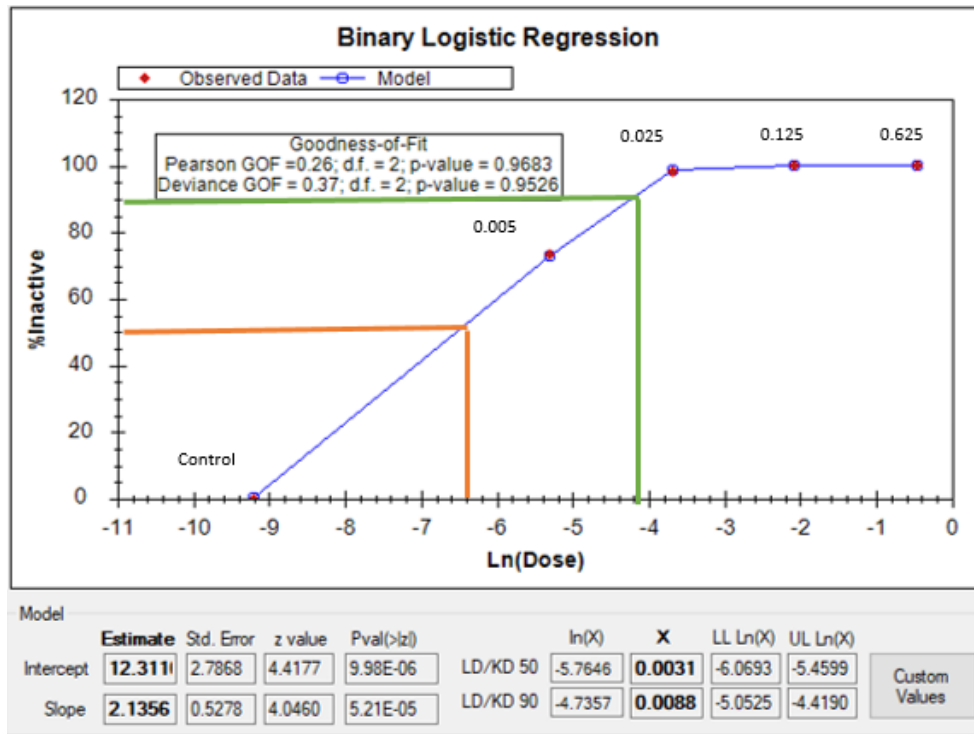


Figure 31. Larvae Resistance Curve for Zone C. LD50 concentration is at 0.0031mg/L while LD90 concentration is at 0.0088 mg/L

## ZONE A, JAN. 16, 2018

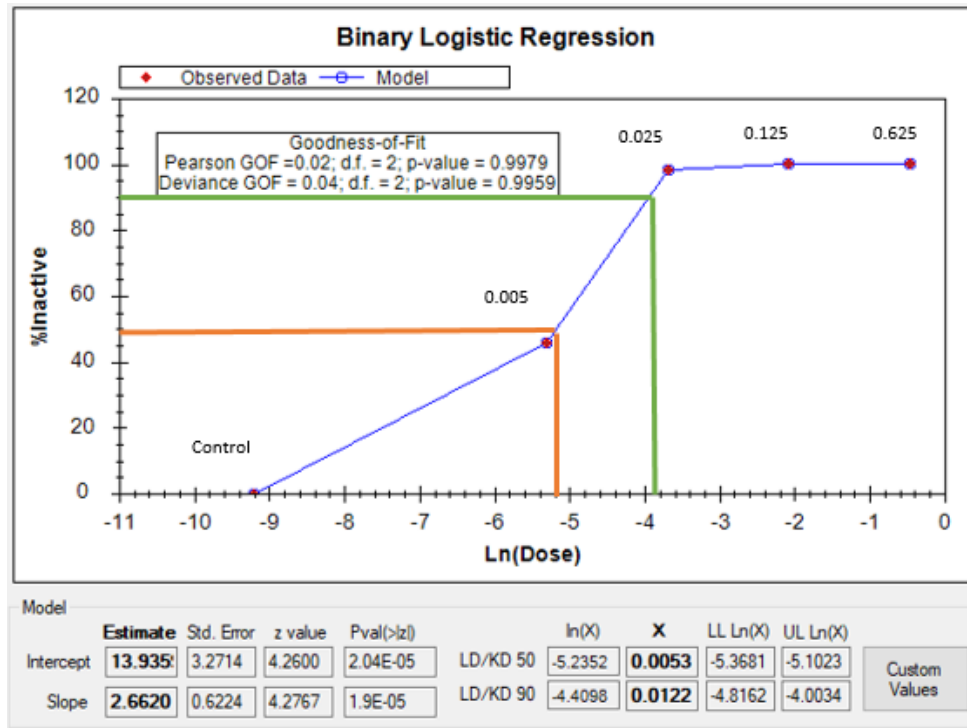


Figure 32. Larvae Resistance Curve for Zone A. LD50 concentration is at 0.0053mg/L while LD90 concentration is at 0.0122 mg/L

**Note:** Sample datasheet used to record resistance data and model used to conduct testing are attached in the appendices on Pages 28-29



## DENGUE ANTIGEN TEST

The adult female *Ae. aegypti* and *Ae. albopictus* mosquitoes that are collected from the BG Sentinel traps in the field are used in a Dengue Antigen Test to detect the presence of Dengue (DENV 1-4) in potentially infected Aedes mosquitoes. The VectorTest® Dengue Assay is a rapid test that identifies the presence or absence of antigens to any of the four Dengue serotypes in mosquitoes. The principle behind the VectorTest® Dengue Assay is the dual monoclonal antibody “sandwich” principle, similar to a pregnancy test, where there is a control zone that will always develop and a test zone that will only develop if there is antigen presence in the mosquito solution.

Prior to processing, all identified Aedes mosquitoes are stored at -20°C. At no point in time are mosquito samples taken out of the fridge unless for processing. For samples of 25 mosquitoes or less, conical grinding tubes/Eppendorf tubes are used for grinding of mosquitoes. For samples of 26 – 50 mosquitoes, plastic culture tubes are used for the grinding of mosquitoes. Once mosquitoes are ground, test strips can be placed in the mosquito suspension and left to react for 20-30 minutes, after which the test results can be read and recorded. All dip sticks with test results are placed in a graphics template and a picture is taken for recording purposes.

ZONE	Total Aedes Adults Captured	WEEK 13 - 16		
		Surveillance Period: January 5-26, 2018		
		Total Aedes Adult <i>Females</i> Tested	Number of Samples run per zone	Positive/Negative for VectorTest®
Date Tested		5 February 2018		
A	4	4	2	NEGATIVE
B	6	3	1	NEGATIVE
C	18	13	4	NEGATIVE
D	6	5	2	NEGATIVE
E	1	1	1	NEGATIVE
F	1	1	1	NEGATIVE
TOTAL			11	0

Table 6. Total adult Aedes mosquitoes captured per zone and total number of female adult Aedes mosquitoes used in Dengue antigen detection test. All zones were negative to the Dengue antigen.

ZONE	Female Adult Aedes for Weeks 13-16 used in VectorTest®				# of Samples Run	# of Positive Samples
	Date of Test: 5 February 2018					
	<i>Ae. aegypti</i>	<i>Ae. albopictus</i>	Aedes Spp. (un-ID)	Total		
A	4	0	0	4	2	0
B	3	0	0	3	1	0
C	10	3	0	13	4	0
D	3	2	0	5	2	0
E	1	0	0	1	1	0
F	1	0	0	1	1	0
TOTAL	22	5	0	27	11	

Table 7. Number of female adult Aedes mosquitoes per zone, sorted by species, used in Dengue Antigen Detection Test

Surveillance Period: Weeks 13 - 16						
DATE OF TEST: 5 February 2018						
BG SENTINEL HOUSE INFORMATION			Number of Female Aedes Adult Mosquitoes Tested in VectorTest®			
ZONE	HOUSE CODE	BG SENTINEL NO.	<i>Ae. aegypti</i>	<i>Ae. albopictus</i>	<i>Aedes Spp (un-ID).</i>	TOTAL/HOUSE
A	BG-H1	1	2	0	0	2
A	BG-H2	2	2	0	0	2
B	BG-H3	3	3	0	0	3
B	BG-H4	4	0	0	0	0
C	BG-H5	5	4	2	0	6
C	BG-H6	6	6	1	0	7
D	BG-H7	7	3	2	0	5
D	BG-H8	8	0	0	0	0
E	BG-H9	9	1	0	0	1
E	BG-H10	10	0	0	0	0
F	BG-H11	11	0	0	0	0
F	BG-H12	12	1	0	0	1
TOTAL			22	5	0	27

Table 8. Number of female adult Aedes mosquitoes per house, sorted by species, used in Dengue Antigen Detection Test.

**NOTE:** Diagram with test results on the dipsticks is attached in Appendices on Page 30

## TRAINING ACTIVITIES

Training sessions are conducted by BVEC staff and International partners to collaborators that work and conduct research in Country. No Training activities we conducted in the month of January 2018

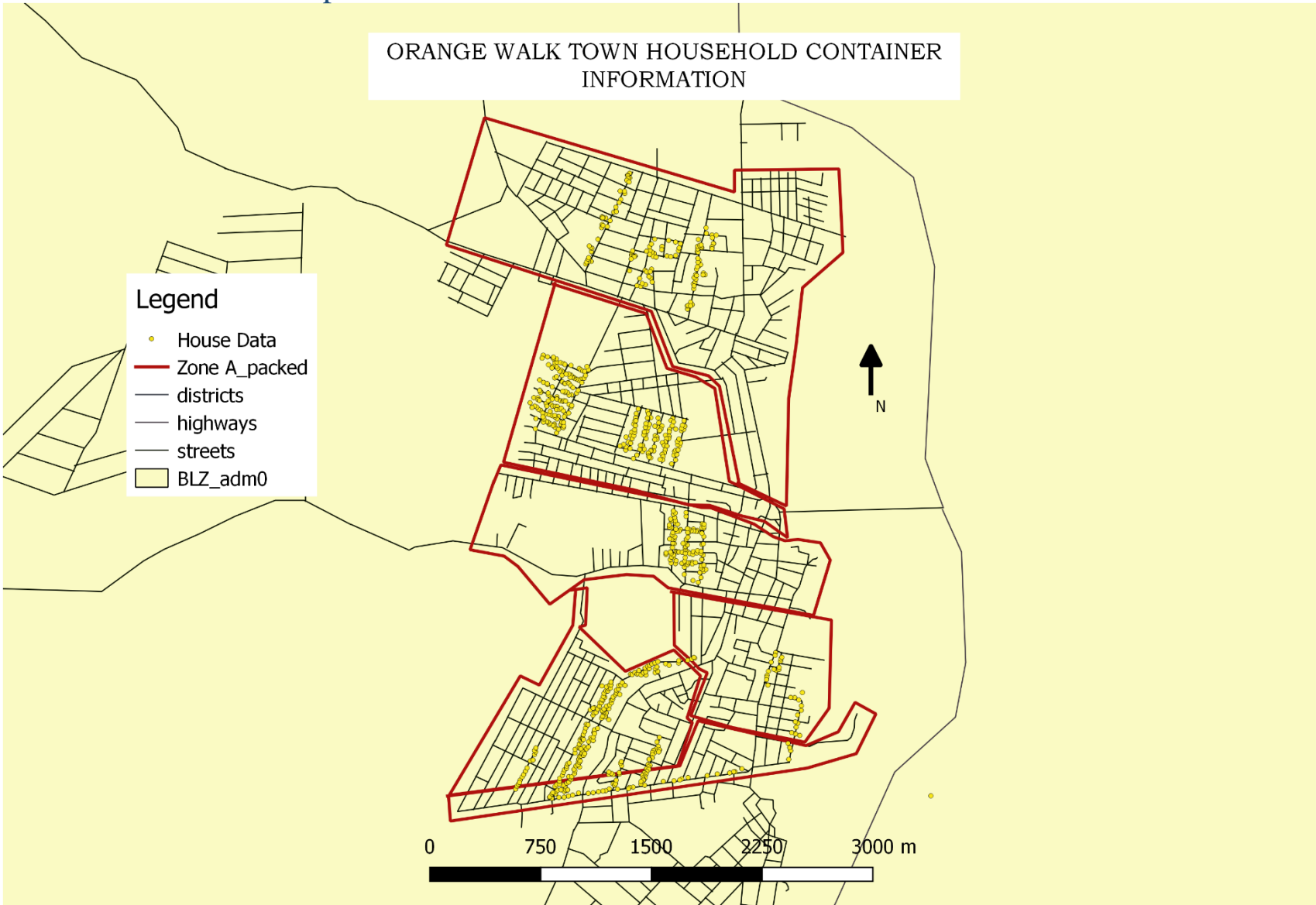
## PRESENTATIONS

Presentations are given by BVEC staff and International partners to collaborators as well as local partners to promote community engagement. Below are a few presentations that have been given so far.

- **Muffles High School Science Fair**
  - Date: January 2016, 2017
  - Venue: Muffles High School
  - Purpose: Science Fair Judging
  - Presenter/s: Donovan Leiva

APPENDICES

APPENDIX I Surveillance Map



Appendix 1. Map of Orange Walk Town divided into zones and displaying points of structures mapped

## APPENDIX II Resistance Datasheet

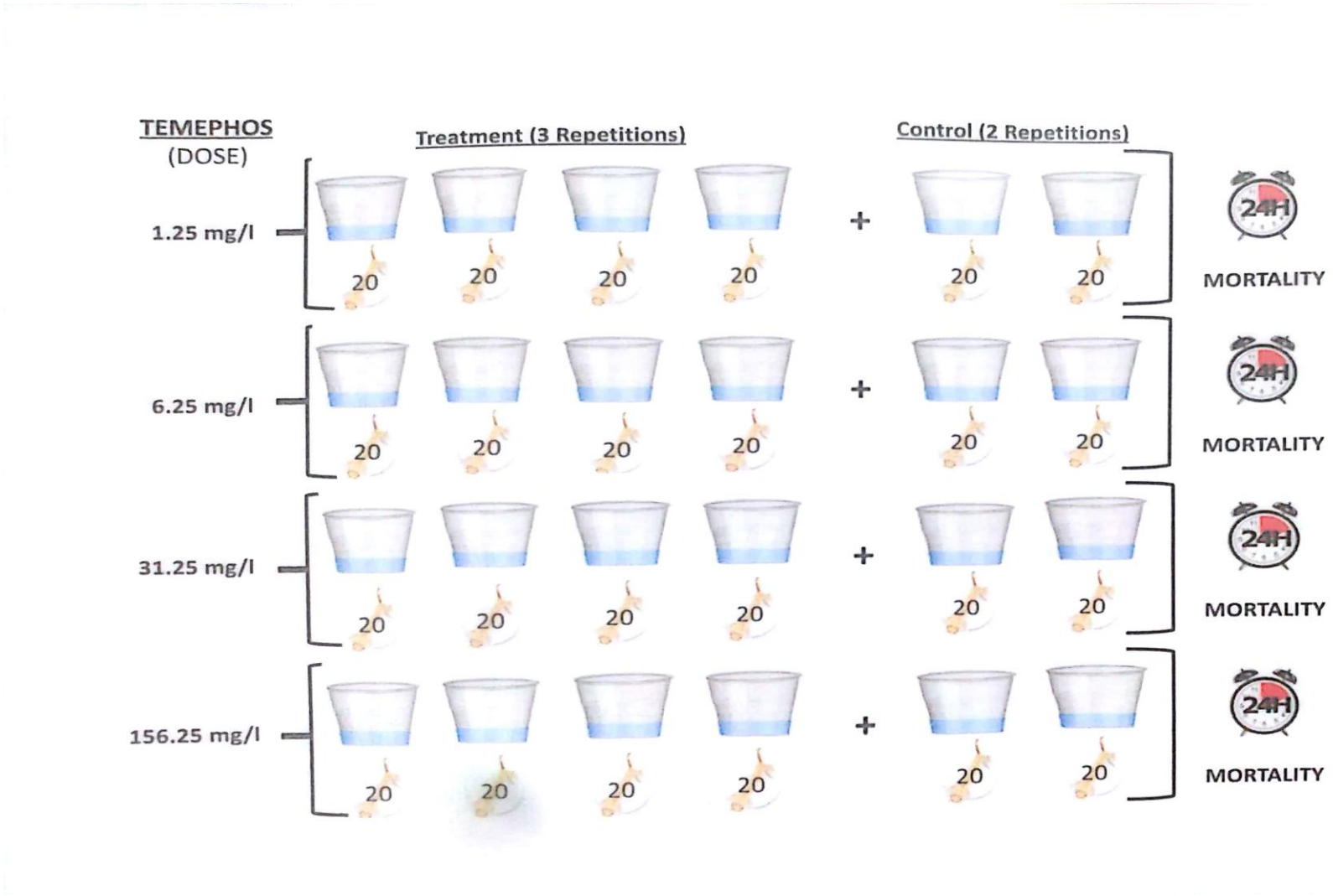
### Small-scale field testing and evaluation of ~~larvicides~~ against mosquito larvae

Experiment No: 2 Starting date: 8/29/17 Location: Orange Walk Town  
 Investigator: \_\_\_\_\_ Assessment date: \_\_\_\_\_ Pre or days posttreatment: \_\_\_\_\_  
 \_\_\_\_\_ Types of Habitat: \_\_\_\_\_ Species Aedes aegypti

Live larvae (L3-4) and pupae (P)/sample							
Treatment ( <del>Temephos</del> )	Sample	Zone: C		Zone: D		Zone:-----	
		24 hrs.	48 hrs.	24 hrs.	48 hrs.	24 hrs.	48 hrs.
Control	1 10	L 10	L 9/ P 1	L 10	L 10		
	2 10	L 10	L 10	L 10	L 10		
	3 10	L 10	L 10	L 10	L 10		
	4 10	L 10	L 10	L 10	L 10		
	5						
	Total 40	40	19	40	40		
	Mean 10	10	9.5	10	10		
	%red						
T1 0.005mg/l	1 10	L 10	L 8/ P 1	L 10	L 9		
	2 10	L 10	L 9	L 10	L 10		
	3 10	L 10	L 9	L 10	L 7		
	4 10	L 10	L 10	L 10	L 8		
	5						
	Total 40	40	36	40	34		
	Mean 10	10	9	10	8.5		
	%red						
T2 0.025mg/l	1 10	L 5	L 2	L 6	L 5		
	2 10	L 6	L 6	L 5	L 5		
	3 10	L 3	L 2	L 2	L 8		
	4 10	L 6	L 1	L 7	L 2		
	5						
	Total 40	20	11	20	20		
	Mean 10	5	2.75	5	5		
	%red						

Appendix 2. Datasheet showing results obtained from a small scale resistance test on Zone C and Zone D in Orange Walk Town. Two concentrations of Temephos were used (0.005mg/L and 0.025mg/L)











APPENDIX III Resistance Protocol



Appendix 3. Model outlining resistance testing protocol


## APPENDIX IV Dengue Test Graphics

Surveillance Period: WKS 13-16 Test Result Graphics | Date: Feb 5, 18

1 2:20pm	2 2:27pm	3 2:30pm	4 2:34pm	5 2:40pm
				
House Code: BG-H1 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H2 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H3 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H5 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H6 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1
6 3:21pm	7 3:32pm	8 3:35pm	9 3:41pm	10 3:48pm
				
House Code: BG-H1 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H9 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H12 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H5 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1	House Code: BG-H6 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 1

Sample 1-8 - *Aedes aegypti* (8)  
Sample 9-11 - *Aedes albopictus* (3)

Surveillance Period: WKS 13-16 Test Result Graphics | Date: Feb 5, 18

11 3:50pm				
				
House Code: BG-H7 Date Collected: WKS 13-16 Researcher's Initials: pmm Sample size: 2	House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:
House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:	House Code: Date Collected: Researcher's Initials:

Appendix 4. Diagram of Vector Test dipsticks displaying results post testing.



## APPENDIX V Consent Form

### Consent for Participation in Research

**Title:** Ovicup/Ovitrap Surveillance for *Ae. aegypti* and *Ae. albopictus* eggs

**Principal Investigator(s):** The Belize Vector and Ecology Center (BVEC)

#### Introduction

The purpose of this form is to provide you with information that may affect your decision as to whether or not to participate in this research study. Read the information below and ask any questions that you might have before deciding whether or not to take part. The person(s) performing this study will answer any of your questions. If you decide to be involved in this study, this form will be used to record your consent.

#### Purpose of the Study

You have been asked to participate in this study by having ovicups/ovitrap set out in your yard for capturing *Ae. aegypti* and *Ae. albopictus* mosquito eggs. The eggs of *Ae. aegypti* and/or *Ae. albopictus* collected in this study will be collected from your yard and hatched in the BVEC laboratory to evaluate mosquito larvicide resistance that may impair the effectiveness of larvicide applications currently being used by the Belize Ministry of Health. This study will also help us to better understand the density of mosquito vectors in the area as well as the abundance of female *Ae. aegypti* and *Ae. albopictus*.

#### What will you be asked to do?

If you agree to participate in this study, you will be asked to provide us with access to your yard so that two ovicups/ovitrap can be placed outside and then retrieved after 7 days of installation. Therefore, ovicups/ovitrap will be out in your yard for an entire week to collect maximum amount of *Ae. aegypti* and/or *Ae. albopictus* eggs. You and individuals at your property will be asked to not move the ovicups/ovitrap from their location and/or disturb them so that the water in the cup or egg paper is not lost. The ovicups/ovitrap will remain in your yard for a total of 6 months. If the study is determined to need to be extended more than 6 months, we will ask if you would like to continue participating.

#### What are the risks involved in this study?

The risks involved with participation in this study are low and may include invasion of privacy. Risk level however, will be reduced by attempting to identify times of the day for ovicup/ovitrap placement and egg paper collection that fit homeowner habits (i.e., time of waking, time of preparing meals, returning from work).

#### How will your privacy and confidentiality be protected if you participate in this research study?

Your privacy and the confidentiality of the data from ovicups/ovitrap placed in your yard will be protected. Household locations will be coded so as not to use family names on data records. The signed consent forms will be stored in a locked, separate location from data records. Your name and any information that would allow you to be identified as an individual will not be used in data presentations, publications or reports resulting from this study.

If it becomes necessary to review study records, information that can be linked to you will be protected to the extent permitted by law. Your research records will not be released without your consent unless required by law or a court order.

#### What are the possible benefits of this study?

You will receive no direct benefit from participating in this study; however, there may be societal benefits as the data collected from this study may be used by the Belize Ministry of Health, Division of Vector Control to guide activities in improving or modifying mosquito larvicide control measures that target *Ae. aegypti* and *Ae. albopictus* mosquitoes that may have the potential to spread diseases such as Zika, dengue and Chikungunya.

**Do you have to participate?**

No, your participation is voluntary. You may decide not to participate at all or, if you start the study, you may withdraw at any time. Withdrawal or refusing to participate will not affect your relationship with the Belize Vector Ecology Center (BVEC) in anyway.

If you would like to participate in this study, sign and return the form to the principal investigator(s). You will receive a copy of this form.

**Will there be any compensation?**

You will not receive any type of payment, compensation or reimbursement for participating in this study.

**Whom to contact with questions about the study?**

Questions can be answered prior to, during or after your participation by contacting BVEC researchers Marla Magaña, Donovan Leiva or Jonathan Kay at 322 -1149 or send an email to [bvec.bz@gmail.com](mailto:bvec.bz@gmail.com).

**Whom to contact with questions concerning your rights as a research participant?**

For questions about your rights or any dissatisfaction with any part of this study, you can contact, anonymously if you wish, BVEC researchers Marla Magaña, Donovan Leiva or Jonathan Kay at 322 - 1149 or send an email to [bvec.bz@gmail.com](mailto:bvec.bz@gmail.com).

**Participation**

If you agree to participate please sign below and return the form to the primary researcher(s).

**Signature**

You have been informed about this study's purpose, procedures, possible benefits and risks, and you have received a copy of this form. You have been given the opportunity to ask questions before you sign, and you have been told that you can ask other questions at any time. You voluntarily agree to participate in this study. By signing this form, you are 18 years or older and you are not waiving any of your legal rights.

---

Printed Name

---

Signature

---

Date

As a representative of this study, I have explained the purpose, procedures, benefits, and the risks involved in this research study.

---

Print Name of Person obtaining consent

---

Signature of Person obtaining consent

---

Date

## APPENDIX VI Glossary

### Glossary of Terms

#### Types of Structures for Mapping Surveillance:

1. **House:** Any household dwelling unit.
2. **House/Store:** Is a household dwelling unit with a small grocery store attached to the structure.
3. **Store:** An independently established grocery store/supermarket/shop.
4. **Apartment/Hotel:** A structure that has rooms or living quarters for rent.
5. **Lab:** Any Medical diagnostic center or laboratory.
6. **Church:** A structure used for public worship.
7. **Bar:** An establishment that is authorized to sell solely alcoholic beverages and light food. May have entertainment such as slot machines and pool tables. Commonly known also as “Cool Spots” or “Lounge” or “Pubs”)
8. **House/Bar:** A structure with the combination of a house and a bar.
9. **Restaurant:** An establishment that is authorized to sell food and drinks. May also sell alcoholic beverages. This also includes small vendor establishments.
10. **School:** An institution that is used for learning and higher education. Either Primary, Secondary or Tertiary level.
11. **Cemetery:** Grounds for which they deceased are laid.
12. **Office:** A government or non-governmental organization office/department. Example: (Immigration, Customs, etc.)
13. **Clinic:** Regional Hospital or any other privately owned establishment where outpatients are given medical treatment or advice, especially of a specialist nature.
14. **Business:** Refers to any other small business establishments such as beauty salons, barbershops, garages, car wash etc.
15. **Other:** Any structure that is distinct from the ones previously named

#### Types of Containers:

1. **Vat/Tank:** A large metallic or plastic container used for storing rain water at homes. Usually collects rain water from gutters that run at the end of roofs.
2. **Drum:** Refers to a metallic or plastic 50 gallon container used for rain water storage. May be used in place of a vat/tank.
3. **Bucket:** A small 1-5 gallon container used to store water (either rain or tap) on a temporary basis.
4. **Bottles/Cans:** Refers to any small empty plastic, glass, aluminum bottles or cans (usually soft-drink or beer) that is found at the premises.
5. **Tires:** Any discarded vehicle tires that do not contain a rim and are able to hold water.
6. **Tree-holes/wells:** Are holes that are natural or man-made in a tree (tree-holes) or in the ground (uncovered wells) that can collect water.
7. **Other:** Any container that is distinct from the ones previously named